

## ANTI-INFLAMMATORY THERAPY IN CYSTIC FIBROSIS

Mario Romano

Department of Biomedical Sciences and Aging Research Center, “G. D’Annunzio”  
University Foundation, Chieti-Pescara, Italy

### *The inflammatory response in Cystic Fibrosis*

An exuberant and persistent acute inflammatory reaction is commonly observed in the lung from cystic fibrosis (CF) patients, representing a key pathogenetic event of lung damage and respiratory insufficiency. CF inflammation is characterized by massive infiltration of activated neutrophils and is sustained by over expression of a repertoire of pro-inflammatory mediators and receptors (1). The mechanisms responsible of lung inflammation in CF are still under debate. One line of evidence suggests that inflammation is triggered by bacterial infection, which develops early in life and is promoted by reduced bacterial clearance, due to mucous viscosity (2). More recent results indicate, instead, that CF airway is prone to inflammation, even in the absence of appreciable bacterial colonization (3). It appears that a dysfunctional Cystic Fibrosis Transmembrane conductance Regulator (CFTR) may trigger over-production of pro-inflammatory cytokines by respiratory epithelial cells either at basal or in response to bacterial infection (4). In vitro studies have shown that inhibition of CFTR activity increases interleukin-8 (IL-8) secretion and NF- $\kappa$ B nuclear translocation in normal human respiratory epithelial cells (5). On the other side, CFTR may regulate functions of circulating cells involved in the immune-surveillance and in the inflammatory response. In particular, polymorphonuclear neutrophils (PMNs) express a functional CFTR protein, which regulates chlorination of bacterial proteins (6). Also, CFTR controls phagosome acidification and bacterial killing in alveolar macrophages (7).

It is becoming clear that, under physiological circumstances, the resolution of the inflammatory response is a well timely-ordered, active process which involves the participation of a number of mediators (8). Thus, pathology may develop also because of alterations in the resolution phase of the inflammatory reaction. Indeed, it has been reported that a defect in generation of the endogenous pro-resolution lipid mediator lipoxin A<sub>4</sub> (LXA<sub>4</sub>) can be observed in CF (9). In aggregate, these observations indicate that a number of factors concur to maintain an acute inflammatory state in CF lung.

This article reviews the anti-inflammatory protocols employed in CF, with the introduction of novel potential experimental approaches.

### *Classic anti-inflammatory drugs*

#### **Steroids**

Although steroids can improve lung function and retard parenchyma deterioration, their use in CF has been undermined by the serious side-effects (cataracts, glucose intolerance, hypertension and growth retardation) associated with long-term administration (10). To minimize such effects, inhaled corticosteroids have been used for both short and long term treatments. Unfortunately, clinical results with aerosolized steroids have been modest (11). Recently, a novel strategy for steroid administration has been proposed. Erythrocytes from patients are loaded with dexamethasone and then reinfused (12). Preliminary data from a small number of patients indicate that erythrocyte-mediated delivery of steroid is relatively safe and is associated with a statistically significant improvement of FEV<sub>1</sub> (12). Larger studies are however required to assess the real efficacy of this treatment to control lung inflammation in CF.

#### **NSAIDs**

High doses of ibuprofen, to reach plasma concentrations between 50 and 100 µg/ml, have been administered to CF children for 4 years in a randomized double-blind trial (13). Initial results showed a slower progression of lung disease in patients treated with ibuprofen (13). A ten-year follow-up of this study showed the persistence of the improvement in lung function in patients taking ibuprofen (14). Furthermore, children with mild lung disease treated with ibuprofen had 33% less reduction in FEV<sub>1</sub> compared with children randomized to placebo (15). The risk of side effects (mainly GI bleeding) and the need for an accurate monitoring of plasma concentrations may limit the use of this drug, although benefits may prevail over the risks. The use of more selective cyclooxygenase-2 (COX-2) inhibitors is not recommended because of their documented cardiovascular side effects in long-term treatments (16).

### *Alternative anti-inflammatory strategies*

#### **Deoxyribonuclease**

DNA released by apoptotic neutrophils infiltrating the CF airways, contributes to mucus viscosity, which in turn promotes bacterial infection and exacerbation of respiratory inflammation. Therefore, treatment with human recombinant deoxyribonuclease (rhDNase) was proposed. 105 CF patients with normal lung function were randomized to receive 2.5 mg/day of nebulized rhDNase or no treatment for three years (17). Untreated patients displayed over time an increase in the number of neutrophils as well as in elastase activity and IL-8 levels in bronchoalveolar lavage fluids (BALF). Such increments were not observed in BALF from patients treated with rhDNase (17). However, no significant variations in FEV<sub>1</sub> were observed between untreated and rhDNase-treated patients. On the other hand, in a 96-week double-blind placebo-controlled randomized study involving a total of 474 CF patients, treatment with dornase alpha reduced the decay in lung functions and the risk of exacerbations (18). Similar

results emerged from the analysis of data from Epidemiologic Survey of Cystic Fibrosis (19). Thus, rhDNAse may exert anti-inflammatory actions in CF patients with mild lung disease, although the impact of this treatment on survival should be better evaluated in longer follow up studies.

### **Leukotriene receptor antagonists**

Leukotrienes (LTs) are generated by 5-Lipoxygenase (LO)-catalyzed conversion of arachidonic acid (20). They are potent regulators of the immune-inflammatory response. In particular, LTB<sub>4</sub>, which is produced by PMNs and macrophages, is a potent chemoattractant for PMNs (21). It activates specific membrane receptors belonging to the G-protein-coupled receptor (GPCR) family and denominated BLT<sub>1</sub> and BLT<sub>2</sub> (22). Increased levels of LTB<sub>4</sub> can be measured in epithelial lining fluid and exhaled breath condensate from CF patients (23,24). Given that BLT<sub>1</sub> plays a key role in the regulation of PMN recruitment in the respiratory tract, contributing to inflammation progression (22), a multi center randomized, double blind, placebo-controlled phase II clinical trial with the BLT<sub>1</sub> antagonist BIIL 284 BS was started. Unfortunately, this trial had to be interrupted because of serious pulmonary adverse effects in patients taking the drug (25).

5-LO generates an additional class of leukotrienes termed cysteinyl leukotrienes (Cys-LT), which regulate vascular tone and permeability and exert pro-inflammatory functions (26). CysLT activate two GPRC receptors, CysLT<sub>1</sub> and CysLT<sub>2</sub> (27). CysLT<sub>1</sub> receptor antagonists are routinely used for treatment of allergic rhinitis and asthma (28). CysLT<sub>1</sub> receptor antagonists have been administered also to CF patients. In one study, montelukast was given to 11 clinically stable CF patients for two weeks (29). Patients displayed some improvement in exercise tolerance (29). In another study, montelukast was administered to 16 patients for three weeks (30). A reduction in levels of eosinophilic cationic protein levels and number of eosinophils was denoted, although functional tests remained unchanged (30). Improvement in respiratory parameters as well as decrease in cough, wheezing, levels of IL-8 and myeloperoxidase in sputum, was instead observed in other clinical investigation with montelukast in CF patients (31). Zafirlukast was also given to thirty CF patients with some improvement in clinical score (32).

Taken together, studies with CysLT<sub>1</sub> inhibitors do not conclusively show a real efficacy of this class of drugs as anti-inflammatory agents in CF.

### ***N*-acetylcysteine**

Oxidant stress may significantly contribute to progression of lung damage in CF. Expression of genes regulating the redox/anti-oxidant systems was altered in the CFTR<sup>ΔF508</sup> mice (33). *N*-acetylcysteine is a sulfhydryl group donor with anti-oxidant properties, which has been largely used as a mucolytic agent. In phase 1 clinical investigation, high doses of *N*-acetylcysteine (0.6 to 1.0 g three times per day) were administered to 18 CF patients for four weeks (34). A significant reduction in sputum elastase activity as well as neutrophil burden was observed, whereas no changes in

pulmonary functions were detected (34). A phase II trial is underway to determine whether, by improving the neutrophil burden, *N*-acetylcysteine can also ameliorate lung function in the long-term.

### **Nitric oxide regulators**

Generation of nitric oxide (NO) by the inducible form of nitric oxide synthase (iNOS) is impaired in CF respiratory epithelial cells (35). This contributes to enhance bacterial infection and to exacerbate airway inflammation. Thus, drugs that increase NO generation and/or bioavailability may be a useful tool to combat inflammation and infection. In this respect, statins, which inhibit the 3-hydroxy-3-methylglutaryl-CoA reductase pathway, restored iNOS expression in CF airway cells, with a mechanism involving the Rho GTPase pathway (36). Statins may also regulate additional anti-inflammatory mechanisms, namely inhibition of cytokine, chemokine production and of chemokine receptor expression (37), NF- $\kappa$ B activation (38), MIP-1 $\alpha$  expression in monocytes (39), inhibition of PMN activation and recruitment (40). The impact of simvastatin administration to CF patients is being currently evaluated.

On the other hand, NO formation may be increased by providing iNOS with L-Arginine, which is enzymatically converted to NO. Administration of inhaled or oral L-Arginine to CF patients resulted in increased NO formation, although a beneficial effect on FEV<sub>1</sub> could not be observed (41).

### **PPAR activators**

Peroxisome proliferators-activated receptors (PPARs) represent a class of transcription factors involved in the regulation of lipid metabolism and of the inflammatory response. Among the three isoforms ( $\alpha$ ,  $\beta$ ,  $\gamma$ ) so far identified, PPAR- $\gamma$  is highly expressed in the respiratory epithelium as well as in alveolar macrophages (42). In addition, a decrease in PPAR- $\gamma$  expression was denoted in the lung from *cfr*<sup>-/-</sup> mice compared to lung from normal mice (43). Activation of PPAR- $\gamma$  by endogenous (15-deoxy- $\Delta$ <sup>12,14</sup>-prostaglandin J<sub>2</sub>) or exogenous (thiozolidinediones) agonists results in down-regulation of pro-inflammatory cytokine expression in airway cells (44) as well as in alveolar macrophages (45). Thus, PPAR- $\gamma$  activators have been proposed for treatment of chronic airway inflammation (46). Together, these observations support the hypothesis that PPAR- $\gamma$  agonists may be beneficial in CF. A clinical trial with pioglitazone in CF patients is underway (47).

### **Cytokines**

Cytokines actively concur to the development and persistence of the inflammatory reaction in CF. In particular, increased production of pro-inflammatory cytokines as well as reduced formation of anti-inflammatory cytokines can be observed in CF patients. Therefore, treatments aimed at regulating the cytokine network may be proposed for CF

therapy. In this respect, IL-10 plays a key role in the resolution of the inflammatory response. Interestingly, IL-10 knockout mice display prolonged inflammation in response to *Pseudomonas aeruginosa* challenge (48). Moreover, treatment with IL-10 improved survival of these mice (49). Therefore, administration of recombinant IL-10 could be a therapeutic option for CF patients. On the other hand, interferon gamma (IFN- $\gamma$ ) can modulate a number of mechanisms of inflammation and alterations in the IFN- $\gamma$  pathway have been detected in CF (50). Inhaled IFN- $\gamma$  was administered to 66 patients randomized to receive 500 or 1000  $\mu$ g IFN- $\gamma$ 1b. Results from this study did not show changes in inflammatory parameters or improvements in respiratory functions (51).

### **Protease inhibitors**

The massive infiltration of PMNs in CF airways is associated with the release of large amounts of proteases stored in PMN granules. These proteases greatly contribute to the development of inflammation and tissue damage. Thus, it has been proposed that supplementation of the airway with anti-proteases may counter-balance the increment in airway protease concentration which occur in CF. Early studies showed that inhaled  $\alpha$ 1-antitrypsin suppressed neutrophil elastase in the airway lining fluid from 12 CF patients (52). In a more recent prospective randomized investigation, 52 CF patients were given 25 mg inhaled  $\alpha$ 1-antitrypsin for four weeks (53). A reduction in pro-inflammatory cytokines and PMNs in the induced sputum was observed, although there were no changes in lung function (53). Aerosol preparation of engineered protein inhibitor human neutrophil elastase (EPI-hNE4) revealed potent inhibitory capability of PMN elastase (54). EPI-hNE4 protected rats from acute lung injury and reduced by 64% PMN influx in rat airway after tracheal instillation of pooled sputum from children with CF (55). Phase II clinical trials of engineered protein inhibitor human neutrophil elastase (EPI-hNE4), which has been shown to efficiently reduce elastase concentrations as well as PMN migration, are being carried out in Europe (47).

### **Antibiotics**

Anti-bacterial therapy may also decrease the inflammation burden in CF airway. A number of evidence indicates that macrolides reduce formation of pro-inflammatory cytokines in vitro and in vivo and also inhibit PMN migration and activation (56). In a multicenter, double-blind, placebo-controlled trial, 87 CF patients were randomized to oral azithromycin (250 or 500 mg) 3 days a week for 168 days, whereas 98 CF patients received placebo. Patients on azithromycin showed a significant improvement in FEV<sub>1</sub> and experienced less exacerbation in comparison with patients on placebo (57). In another study, 27 CF patients were treated with oral clarithromycin (250 mg every other day) for 12 months (58). A reduction in TNF- $\alpha$  and IL-8 levels in plasma and sputum associated with a significant amelioration of lung functions was observed (58).

## Stop signals and pro-resolution mediators of inflammation

The inflammatory response is a well ordered sequelae of events that physiologically culminates with resolution and restitution at integrum. It is now clear that the resolution phase of inflammation is an active process, tightly regulated by a wide number of mediators. Thus, the understanding of mechanisms involved in resolution, may provide the basis for alternative anti-inflammatory treatments.

Metabolism of unsaturated fatty acids generates a number of pro-resolution compounds. In particular, arachidonic acid (AA) is converted by cooperation between lipoxygenases into lipoxins (LX), whereas aspirin-treated cyclooxygenase 2 (COX-2) in cooperation with 5-LO yields the 15-epi-LX (59). On the other side, eicosapentaenoic acid (EPA) is converted by aspirin-acetylated COX-2 into resolvins of the E-series (RvE1), whereas docosahexanoic acid (DHA) can give rise to protectins/neuroprotectins (PD1) or 17S-resolvins of the D-series (RvD1) when attacked by LO or 17R-resolvins of the D-series (AT-RvD1) in the presence of aspirin-acetylated COX-2 (60). LX and epi-LX as well as resolvins and protectins exert potent anti-inflammatory bioactions in vitro and in vivo (59,60). Interestingly, BALF from CF patients display decreased lipoxin A<sub>4</sub> (LX<sub>4</sub>) levels (61). In addition, a LXA<sub>4</sub> stable analog suppressed neutrophilic inflammation and reduced disease severity in a mouse model of CF (61). On the other hand, a deficiency in DHA, which is the precursor of protectins and resolvins, has been observed in CF (62), providing a rationale for DHA trials in CF (63). Administration to 5 CF patients of 70 mg of DHA/kg body weight/d for six weeks, determined a significant increase in tissue DHA levels (64). Long-term administration of DHA to *Cftr*<sup>-/-</sup> mice resulted in a significant improvement of inflammatory liver disease (65).

These observations indicate that alterations in endogenous pro-resolution signals may be also involved in CF inflammation. It may be therefore hypothesized that administration of pro-resolution drugs may represent an alternative therapeutic option to combat inflammation in CF.

### *Concluding remarks*

It is now clear that controlling inflammation is a challenging, however crucial, task in CF therapy. A variety of approaches have been experimented with mixed results (Table 1). It is essential to decode the key and peculiar circuits of CF inflammation to establish a successful treatment, since conventional anti-inflammatory drugs have shown limited therapeutic efficacy. If, on one side, there is a need for more, prospective clinical trials, on the other side the indications provided by basic science should be translated into novel approaches. In this respect, the synthesis and the use of pro-resolutions drugs may be worth pursuing.

## REFERENCES

1. Koehler DR, Downey GP, Sweezey NB, Tanswell AK, Hu J. Lung inflammation as a therapeutic target in cystic fibrosis. 2004 *Am J Respir Cell Mol Biol.* 31:377-381.
2. Davis PB, Drumm ML, Konstan MW. State of the Art: Cystic Fibrosis. 1996 *Am J Resp Crit Care Med.* 154:1229-1256.
3. Carrabino S, Carpani D, Livraghi A, Di Cicco M, Costantini G, Copreni E, Colombo C, Conese M. Dysregulated interleukin-8 secretion and NF-kappaB activity in human cystic fibrosis nasal epithelial cells. 2006 *J Cyst Fibros.* 5:113-119.
4. Stecenko AA, King G, Torii K, Breyer RM, Dworski R, Blackwell TS, Christman JW, Brigham KL. Dysregulated cytokine production in human cystic fibrosis bronchial epithelial cells. 2001 *Inflammation* 25:145-155.
5. Perez A, Issler AC, Cotton CU, Kelley TJ, Verkman AS, Davis PB. CFTR inhibition mimics the cystic fibrosis inflammatory profile. 2006 *Am J Physiol Lung Cell Mol Physiol.* Aug 18; [Epub ahead of print].
6. Painter RG, Valentine VG, Lanson NA Jr, Leidal K, Zhang Q, Lombard G, Thompson C, Viswanathan A, Nauseef WM, Wang G, Wang G. CFTR Expression in human neutrophils and the phagolysosomal chlorination defect in cystic fibrosis. 2006 *Biochemistry*45:10260-10269.
7. Di A, Brown ME, Deriy LV, Li C, Szeto FL, Chen Y, Huang P, Tong J, Naren AP, Bindokas V, Palfrey HC, Nelson DJ. CFTR regulates phagosome acidification in macrophages and alters bactericidal activity. 2006 *Nat Cell Biol.* 8:933-944.
8. Serhan CN, Brain SD, Buckley CD, Gilroy DW, Haslett C, O'Neill LA, Perretti M, Rossi AG, Wallace JL. Resolution of inflammation: state of the art, definitions and terms. 2007 *FASEB J.* 21:325-332.
9. Karp CL, Flick LM, Park KW, Softic S, Greer TM, Keledjian R, Yang R, Uddin J, Guggino WB, Atabani SF, Belkaid Y, Xu Y, Whitsett JA, Accurso FJ, Wills-Karp M, Petasis NA. Defective lipoxin-mediated anti-inflammatory activity in the cystic fibrosis airway. 2004 *Nat Immunol.* 5:388-392.
10. Cheng K, Ashby D, Smyth R. Oral steroids for cystic fibrosis. 2000 *Cochrane Database Syst Rev.* 2:CD000407.
11. Dezateux C, Walters S, Balfour-Lynn I. Inhaled corticosteroids for cystic fibrosis [Cochrane methodology review]. 2003 In: *The Cochrane Library* issue 4. Chichester (UK): John Wiley & Sons, Ltd.
12. Lucidi V, Tozzi AE, Bella S, Turchetta A. A pilot trial on safety and efficacy of erythrocyte-mediated steroid treatment in CF patients. 2006 *BMC Pediatr.* 6:17.
13. Konstan MW, Byard PJ, Hoppel CL, Davis PB. Effect of high-dose ibuprofen in patients with cystic fibrosis. 1995 *N Engl J Med.* 332:848-844.
14. Konstan MW, Schluchter MD, Storfer-Isser, Davis PB. Use of ibuprofen for the treatment of airway inflammation in CF: an update. 2002 *Pediatr Pulmonol. Suppl* 24:164-165.

15. Schluchter MD, Konstan MW, Xue L, Davis PB. Relationship between high-dose ibuprofen use and rate of decline in FEV1 among young patients with mild lung disease in the CFF Registry. 2004 *Pediatr Pulmonol. Suppl* 27:A385, 322.
16. Rodríguez LA, Patrignani P. The ever growing story of cyclo-oxygenase inhibition. 2006 *Lancet* 368:1745-1747.
17. Paul K, Rietschel E, Ballmann M, Griese M, Worlitzsch D, Shute J, Chen C, Schink T, Döring G, van Koningsbruggen S, Wahn U, Ratjen F; Bronchoalveolar Lavage for the Evaluation of Antiinflammatory Treatment Study Group. Effect of treatment with dornase alpha on airway inflammation in patients with cystic fibrosis. 2004 *Am J Respir Crit Care Med.* 169:719-725.
18. Quan JM, Tiddens HA, Sy JP, McKenzie SG, Montgomery MD, Robinson PJ, Wohl ME, Konstan MW; Pulmozyme Early Intervention Trial Study Group. A two-year randomized, placebo-controlled trial of dornase alfa in young patients with cystic fibrosis with mild lung function abnormalities. 2001 *J Pediatr.* 139:813-820.
19. Konstan MW, Wagener JS, Pasta DJ, et al. Pulmozyme (dornase alfa) use is associated with a slower rate of lung function decline in patients with cystic fibrosis. 2006 *Pediatr Pulmonol. Suppl* 29:A370, 337.
20. Romano M, Claria J. Cyclooxygenase-2 and 5-lipoxygenase converging functions on cell proliferation and tumor angiogenesis: implications for cancer therapy. 2003 *FASEB J.* 17:1986-1995.
21. Funk, CD. Prostaglandins and leukotrienes: advances in eicosanoid biology. 2001 *Science* 294:1871-1875.
22. Okuno T, Yokomizo T, Hori T, Miyano M, Shimizu T. Leukotriene B<sub>4</sub> receptor and the function of its helix 8. 2005 *J Biol Chem.* 280:32049-32052.
23. Konstan MW, Walenga RW, Hilliard KA, Hilliard JB. Leukotriene B<sub>4</sub> markedly elevated in the epithelial lining fluid of patients with cystic fibrosis. 1993 *Am Rev Respir Dis.* 148:896-901.
24. Carpagnano GE, Barnes PJ, Geddes DM, Hodson ME, Kharitonov SA. Increased Leukotriene B<sub>4</sub> and Interleukin-6 in Exhaled Breath Condensate in Cystic Fibrosis. 2003 *Am J Respir Crit Care Med.* 167:1109-1112.
25. Konstan MW, Doring G, Lands LC, Hilliard KA, Koker P, Bhattacharya B, Staab A, Hamilton AL on behalf of the Investigators and Coordinators of BI Trial 543.45. Results of a phase II clinical trial of BIIL 284 BS (a LTB<sub>4</sub> receptor antagonist) for the treatment of CF lung disease. 2005 *Pediatr Pulmonol. Suppl* 28:125-7.
26. Peters-Golden M, Canetti C, Mancuso P, Coffey MJ. Leukotrienes: underappreciated mediators of innate immune responses. 2005 *J Immunol.* 174:589-594.
27. Evans JF. The cysteinyl leukotriene receptors. 2003 *Prostaglandins Leukot Essent Fatty Acids.* 69:117-122.
28. Capra V, Ambrosio M, Riccioni G, Rovati GE. Cysteinyl-leukotriene receptor antagonists: present situation and future opportunities. 2006 *Curr Med Chem.* 13:3213-3226.
29. Morice AH, Kastelik JA, Aziz I. Montelukast sodium in cystic fibrosis. 2001 *Thorax* 56:244-245.



30. Schmitt-Grohé S, Eickmeier O, Schubert R, Bez C, Zielen S. Anti-inflammatory effects of montelukast in mild cystic fibrosis. 2002 *Ann Allergy Asthma Immunol.* 89:599-605.
31. Stelmach I, Korzeniewska A, Stelmach W, Majak P, Grzelewski T, Jerzynska J. Effects of montelukast treatment on clinical and inflammatory variables in patients with cystic fibrosis. 2005 *Ann Allergy Asthma Immunol.* 95:372-380.
32. Conway SP, Etherington C, Peckham DG, Whitehead A. A pilot study of zafirlukast as an anti-inflammatory agent in the treatment of adults with cystic fibrosis. 2003 *J Cyst Fibros.* 2:25-28.
33. Xu Y, Liu C, Clark JC, Whitsett JA. Functional genomic responses to cystic fibrosis transmembrane conductance regulator (CFTR) and CFTR<sup>Δ508</sup> in the lung. 2006 *J Biol Chem.* 281:11279-11291.
34. Tirouvanziam R, Conrad CK, Bottiglieri T, Herzenberg LA, Moss RB, Herzenberg LA. High-dose oral N-acetylcysteine, a glutathione prodrug, modulates inflammation in cystic fibrosis. 2006 *Proc Natl Acad Sci U S A.* 103:4628-4633.
35. Zheng S, De BP, Choudhary S, Comhair SA, Goggans T, Slee R, Williams BR, Pilewski J, Haque SJ, and Erzurum SC. Impaired innate host defense causes susceptibility to respiratory virus infections in cystic fibrosis. 2003 *Immunity* 18: 619-630.
36. Kraynack NC, Corey DA, Elmer HL, Kelley TJ. Mechanisms of NOS2 regulation by Rho GTPase signaling in airway epithelial cells. 2002 *Am J Physiol Lung Cell Mol Physiol* 283:L604-L611.
37. Yin R, Zhu J, Shao H, Cheng X, Feng X, Li Z, Jing H. Inhibition of chemokine receptor CCR2 and CCR5 expression contributes to simvastatin-induced attenuation of cardiac allograft vasculopathy. 2007 *J Heart Lung Transplant.* 26:485-493.
38. Lee JY, Kim JS, Kim JM, Kim N, Jung HC, Song IS. Simvastatin inhibits NF-κB signaling in intestinal epithelial cells and ameliorates acute murine colitis. 2007 *Int Immunopharmacol* 7:241-248.
39. Bruegel M, Teupser D, Haffner I, Mueller M, Thiery J. Statins reduce macrophage inflammatory protein-1α expression in human activated monocytes. 2006 *Clin Exp Pharmacol Physiol.* 33:1144-1149.
40. Fischetti F, Carretta R, Borotto G, Durigutto P, Bulla R, Meroni PL, Tedesco F. Fluvastatin treatment inhibits leucocyte adhesion and extravasation in models of complement-mediated acute inflammation. 2004 *Clin Exp Immunol.* 135:186-193.
41. Grasemann H, Grasemann C, Kurtz F, Tietze-Schillings G, Vester U, Ratjen F. Oral L-arginine supplementation in cystic fibrosis patients: a placebo-controlled study. 2005 *Eur Respir J.* 25:62-68.
42. Hsun-Wei Huang T, Razmovski-Naumovski V, Kota BP, Shu-Hsuan Lin D, Roufogalis BD. The pathophysiological function of peroxisome proliferator-activated receptor-γ in lung-related diseases. 2005 *Respir Res.* 6:102.
43. Ollero M, Junaidi O, Zaman MM, Tzamelis I, Ferrando AA, Andersson C, Blanco PG, Bialecki E, Freedman SD. Decreased expression of peroxisome proliferator activated receptor gamma in cftr<sup>-/-</sup> mice. 2004 *J Cell Physiol.* 200:235-244.

44. Wang AC, Dai X, Luu B, Conrad DJ. Peroxisome proliferators-activated receptor-gamma regulates airway epithelial cell activation. 2001 *Am. J. Respir Cell Mol Biol.* 24:688-693.
45. Asada, K., Sasaki, S., Suda, T., Chida, K., Nakamura, H., 2003. Antiinflammatory roles of peroxisome proliferator-activated receptor gamma in human alveolar macrophages. 2003 *Am J Respir Crit Care Med.* 169:195-200.
46. Belvisi MG, Hele DJ, Birrell MA. Peroxisome proliferator-activated receptor gamma agonists as therapy for chronic airway inflammation. 2006 *Eur J Pharmacol.* 533:101-109.
47. Chmiel JF, Konstan MW. Inflammation and anti-inflammatory therapies for cystic fibrosis. 2007 *Clin Chest Med* 28:331-346.
48. Chmiel JF, Konstan MW, Saadane A, Krenicky JE, Lester Kirchner H, Berger M. Prolonged inflammatory response to acute *Pseudomonas* challenge in interleukin-10 knockout mice. 2002 *Am J Respir Crit Care Med.* 165:1176-1181.
49. Chmiel JF, Konstan MW, Knesebeck JE, Hilliard JB, Bonfield TL, Dawson DV, Berger M. IL-10 attenuates excessive inflammation in chronic *Pseudomonas* infection in mice. 1999 *Am J Respir Crit Care Med.* 160:2040-2047
50. Kelley TJ, Elmer HL. In vivo alterations of IFN regulatory factor-1 and PIAS1 protein levels in cystic fibrosis epithelium. 2000 *J Clin Invest.* 106:403-410.
51. Moss RB, Mayer-Hamblett N, Wagener J, Daines C, Hale K, Ahrens R, Gibson RL, Anderson P, Retsch-Bogart G, Nasr SZ, Noth I, Waltz D, Zeitlin P, Ramsey B, Starko K. Randomized, double-blind, placebo-controlled, dose-escalating study of aerosolized interferon gamma-1b in patients with mild to moderate cystic fibrosis lung disease. 2005 *Pediatr Pulmonol.* 39:209-218.
52. McElvaney NG, Hubbard RC, Birrer P, Chernick MS, Caplan DB, Frank MM, Crystal RG. Aerosol alpha 1-antitrypsin treatment for cystic fibrosis. 1991 *Lancet* 337:392-394.
53. Griese M, Latzin P, Kappler M, Weckerle K, Heinzlmaier T, Bernhardt T, Hartl D. alpha1-Antitrypsin inhalation reduces airway inflammation in cystic fibrosis patients. 2007 *Eur Respir J.* 29:240-250.
54. Grimbert D, Vecellio L, Delepine P, Attucci S, Boissinot E, Poncin A, Gauthier F, Valat C, Saudubray F, Antonioz P, Diot P. Characteristics of EPI-hNE4 aerosol: a new elastase inhibitor for treatment of cystic fibrosis. 2003 *J Aerosol Med.* 16:121-129.
55. Delacourt C, Herigault S, Delclaux C, Poncin A, Levame M, Harf A, Saudubray F, Lafuma C. Protection against acute lung injury by intravenous or intratracheal pretreatment with EPI-HNE-4, a new potent neutrophil elastase inhibitor. 2002 *Am J Respir Cell Mol Biol.* 26:290-297.
56. Ianaro A, Ialenti A, Maffia P, Sautebin L, Rombolà L, Carnuccio R, Iuvone T, D'Acquisto F, Di Rosa M. Anti-inflammatory activity of macrolide antibiotics. 2000 *J Pharmacol Exp Ther.* 292:156-163.
57. Saiman L, Marshall BC, Mayer-Hamblett N, Burns JL, Quittner AL, Cibene DA, Coquette S, Fieberg AY, Accurso FJ, Campbell PW; Macrolide Study Group. Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*. 2003 *JAMA* 290:1749-1756.

58. Pukhalsky AL, Shmarina GV, Kapranov NI, Kokarovtseva SN, Pukhalskaya D Kashirskaja NJ. Anti-inflammatory and immunomodulating effects of clarithromycin in patients with cystic fibrosis lung disease. 2004 *Mediators Inflamm.* 13:111-117.
59. Romano M. Lipid mediators: Lipoxin and aspirin-triggered 15-epi-lipoxins. 2006 *Inflamm Allergy Drug Targets* 5:81-90.
60. Ariel A, Serhan CN. Resolvins and protectins in the termination program of acute inflammation 2007 *Trends Immunol.* 28:176-183.
61. Karp CL, Flick LM, Park KW, Softic S, Greer TM, Keledjian R, Yang R, Uddin J, Guggino WB, Atabani SF, Belkaid Y, Xu Y, Whitsett JA, Accurso FJ, Wills-Karp M, Petasis NA. Defective lipoxin-mediated anti-inflammatory activity in the cystic fibrosis airway. 2004 *Nat Immunol.* 5:388-392.
62. Freedman SD, Katz MH, Parker EM, Laposata M, Urman MY, Alvarez JG. A membrane lipid imbalance plays a role in the phenotypic expression of cystic fibrosis in CFTR *-/-* mice. 1999 *Proc Natl Acad Sci.* 96:13995-4000.
63. Van Biervliet S, Van Biervliet JP, Robberecht E, Christophe A. Docosahexaenoic acid trials in cystic fibrosis: A review of the rationale behind the clinical trials. 2005 *J Cyst Fibros* 4:27-34.
64. Jumpsen JA, Brown NE, Thomson AB, Paul Man SF, Goh YK, Ma D, Clandinin MT. Fatty acids in blood and intestine following docosahexaenoic acid supplementation in adults with cystic fibrosis. 2006 *J Cyst Fibros.* 5:77-84.
65. Beharry S, Ackerley C, Corey M, Kent G, Heng Y-M, Christensen H, Luk C, Yantiss RK, Nasser IA, Zaman M, Freedman SD, Durie PR. Long-term docosahexaenoic acid therapy in a congenic murine model of cystic fibrosis. 2007 *Am J Physiol Gastrointest Liver Physiol* 292: G839-G848.

TABLE 1

ANTI-INFLAMMATORY DRUGS IN CF			
Drug	Administration route	Results and Remarks	Reference
STEROIDS	Oral Inhaled Erythrocyte	Improvement in lung function. Serious adverse effects in long-term treatment Insufficient evidence of benefit or risk. Small populations studied. Some improvement in FEV <sub>1</sub> . Evidence is too preliminary.	[10] [11] [12]
NSAIDs			
<i>Ibuprofen</i>	Oral	Consistent slow progression of lung disease. Modest risk of GI bleeding.	[13-15]
<i>Selective COX-2 inhibitors</i>		No data available in CF. Cardiovascular risk in long-term administration.	[16]
DEOXYRIBONUCLEASE	Inhaled	Reduction in inflammatory markers in BALF. Reduction in decay of lung functions and risk of exacerbations. Longer follow up trials are needed.	[17-19]
LEUKOTRIENE-R ANTAGONISTS			
<i>BLT<sub>1</sub></i>	Oral	Trial stopped because of serious respiratory adverse effects.	[25]
<i>CysLT<sub>1</sub></i>	Oral	Modest improvement in exercise tolerance, respiratory symptoms and inflammatory parameters. No conclusive evidence of benefit.	[29-32]
ANTI-OXIDANTS			
<i>N-acetylcysteine</i>	Oral	Significant reduction in sputum elastase activity and neutrophil burden. No changes in pulmonary functions. More consistent evidence needed. Phase II	[34]

		trial is underway.	
NITRIC OXIDE REGULATORS			
<i>Statins</i>	Oral	Under evaluation.	
<i>L-Arginine</i>	Oral or Inhaled	Increased NO formation. No changes in FEV <sub>1</sub> .	[41]
PPAR ACTIVATORS			
PPAR- $\gamma$	Oral	Down-regulation of pro-inflammatory cytokine expression in airway cells and in alveolar macrophages. Under clinical evaluation.	[44-45]
CYTOKINES			
IFN- $\gamma$	Inhaled	No changes in respiratory functions or inflammation markers.	[51]
IL-10		Prolonged inflammation in response to <i>Pseudomonas aeruginosa</i> in IL-10 knockout mice. IL-10 improved survival of IL-10 knockout mice.	[48-49]
PROTEASE INHIBITORS			
<i><math>\alpha</math>1-antitrypsin</i>	Inhaled	Suppression of neutrophil elastase. Reduction in pro-inflammatory cytokines and PMNs in the induced sputum. NO changes in lung function.	[52-53]
<i>EPI-hNE4</i>	Inhaled	Protection of rats from acute lung injury. Reduction of PMN influx in rat airway after tracheal instillation of pooled sputum from children with CF. Phase II clinical trial is underway in Europe.	[54-55]
ANTIBIOTICS			

<i>Azithromycin</i>	Oral	Significant improvement in FEV <sub>1</sub> . Less exacerbations.	[57]
<i>Clarithromycin</i>	Oral	Reduction in TNF- $\alpha$ and IL-8 levels in plasma and sputum. Significant improvement of lung functions.	[58]
PRO-RESOLUTION STOP SIGNALS			
<i>Lipoxins</i>	Oral	Suppression of neutrophilic inflammation and reduction disease severity in a mouse model of CF.	[61]
<i>Protectins</i>		Potent anti-inflammatory activity in vitro and in animal models. No data available in CF.	[62]
<i>Resolvins</i>		Potent anti-inflammatory activity in vitro and in animal models. No data available in CF.	[62]
<i>DHA</i>	Oral	Increase in tissue DHA levels in CF patients. Significant improvement of liver disease in <i>Cftr</i> <sup>-/-</sup> mice.	[64-65]