

# Il futuro del genome editing

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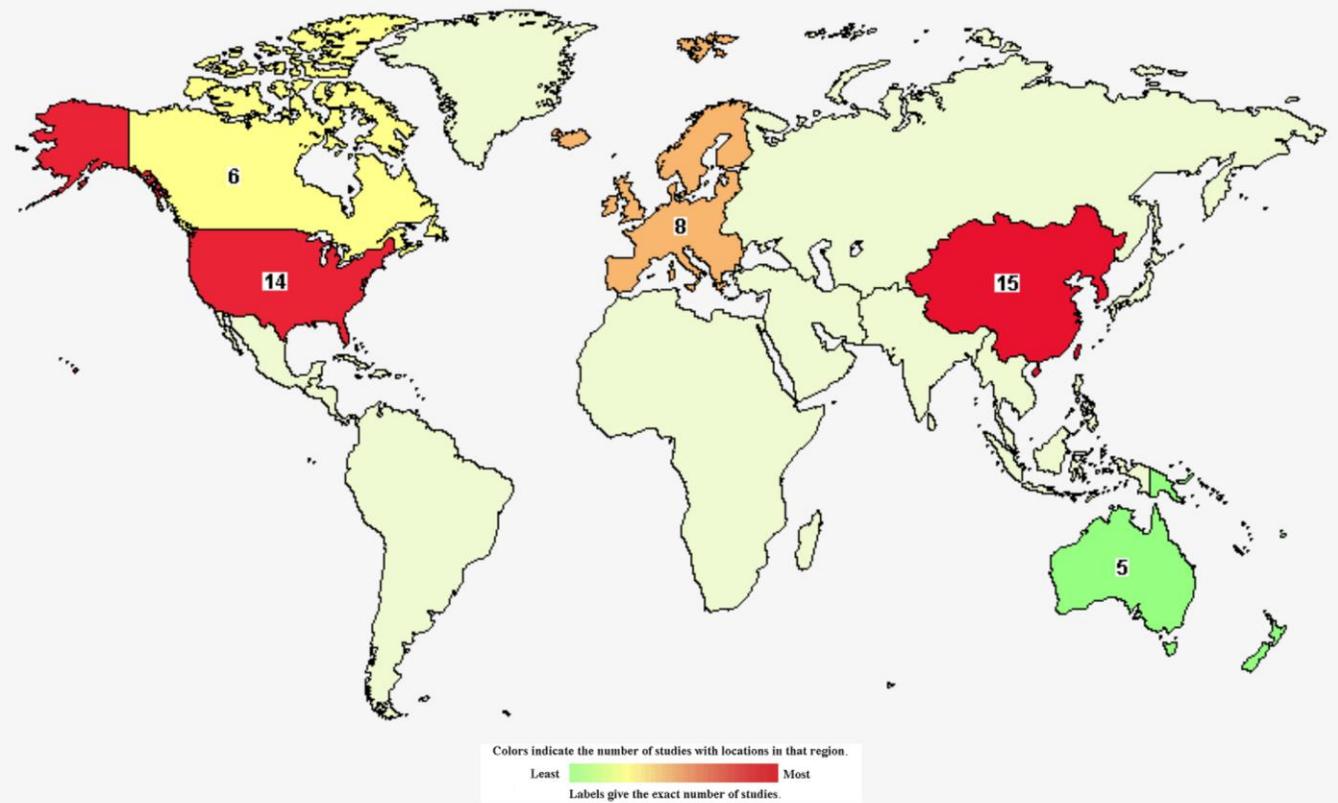
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# World map clinical trials using CRISPR-Cas9 technology

[A similar map is available for all studies in ClinicalTrials.gov](#)

Click on the map below to show a more detailed map (when available) or search for studies (when map not available).



Source: <https://ClinicalTrials.gov>

## Selected trials from the world list:

### Europe:

- Beta-Thalassemia
- Sickle Cell Disease

### US:

- Edited T cells (TCR and PD1 ko)
- Multiple Myeloma
- Melanoma
- Synovial Sarcoma
- Myxoid/Round Cell Liposarcoma

- Blindness (Leber Congenital Amaurosis)

### China

- CAR-T cells (CD19, CD20, CD22)
- B Cell Leukemia
- B Cell Lymphoma

- CCR5 ko HIV-1-infection

- PD1 ko for EBV (Epstein Barr) malignancies (Gastric, Nasopharyngeal, Lymphoma)

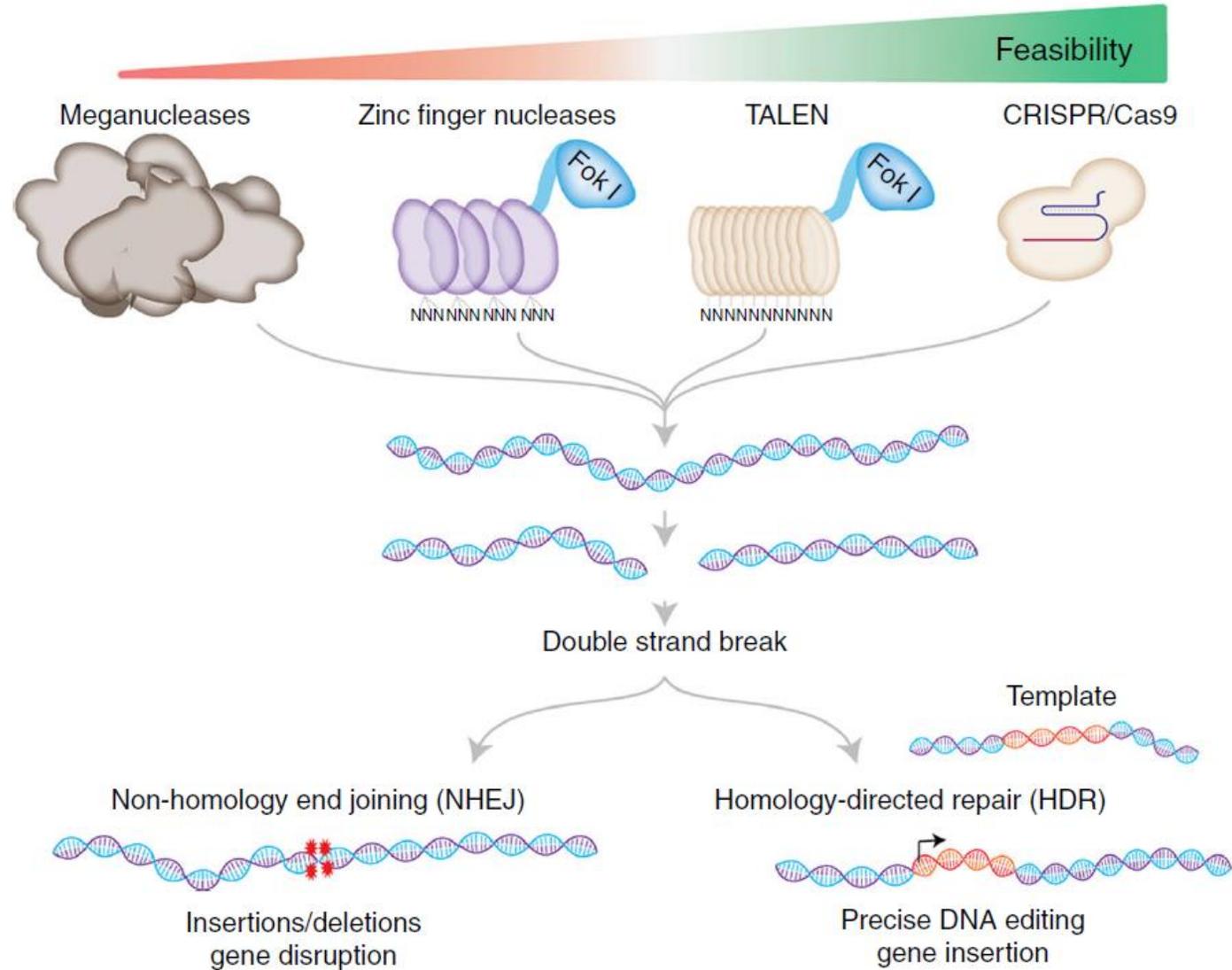
## I. Background

- Avanzamenti e ostacoli del genome editing
- Proprietà in-vitro/in-vivo del genome editing
- Aggiornamenti di genome editing per la fibrosi cistica

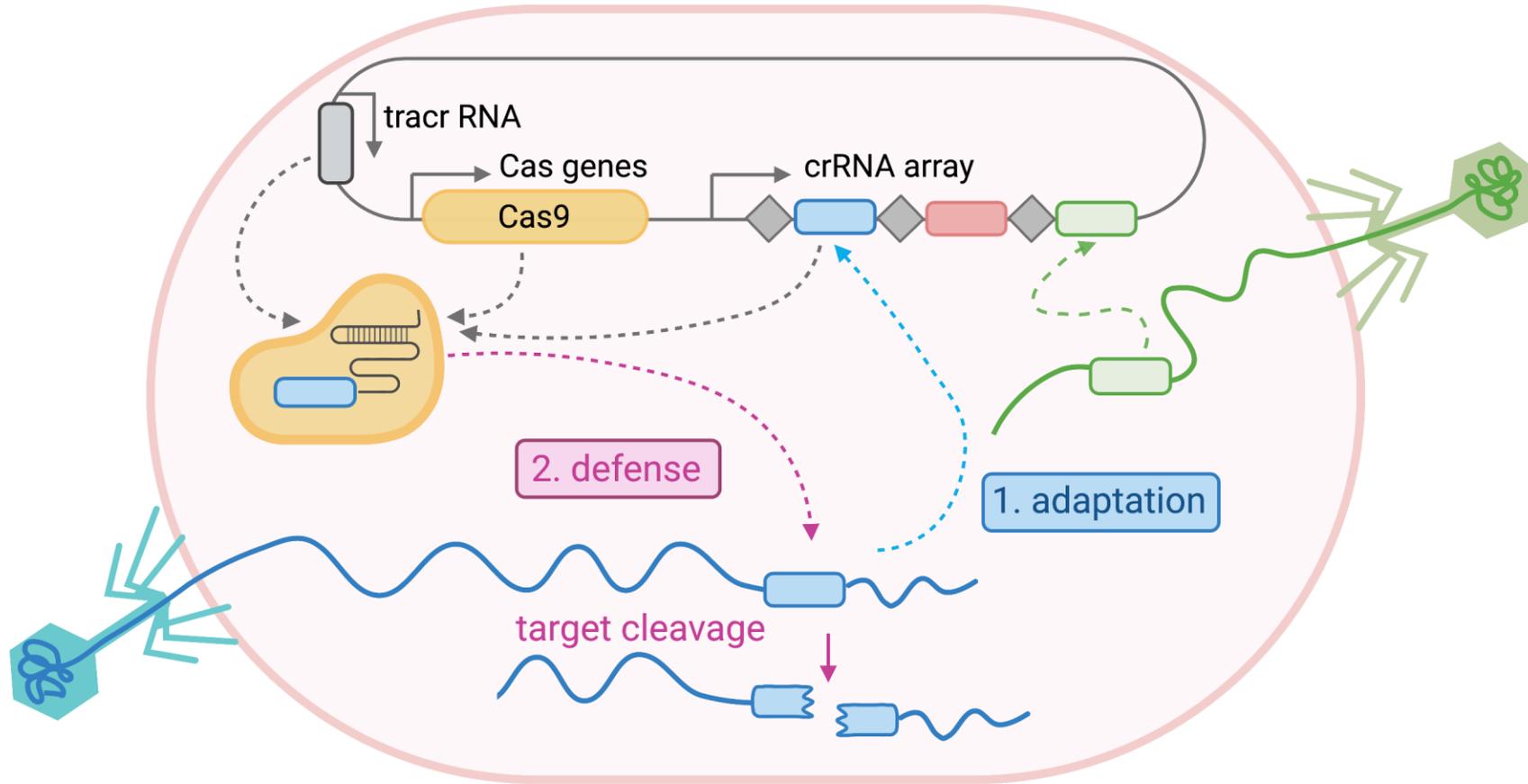
## II. Esempio di correzione di mutazione di splicing

- Correzione della mutazione di splicing 3272-26A>G utilizzando CRISPR-Cas

# Genome editing technology

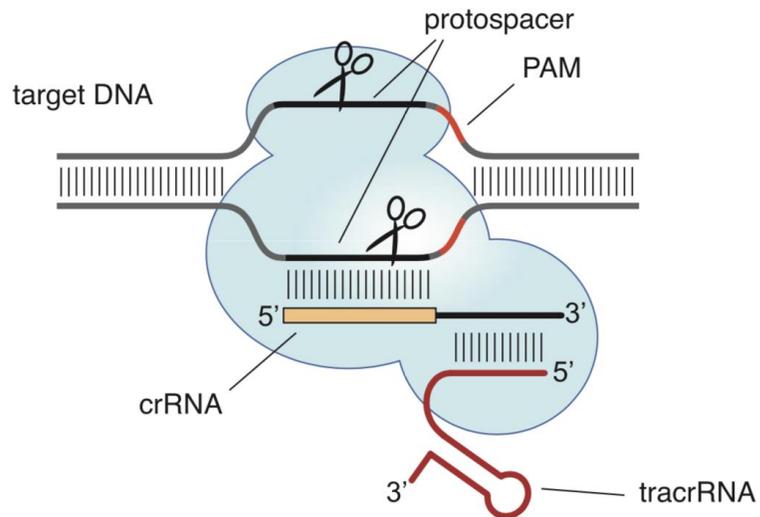


# Host defense in bacteria

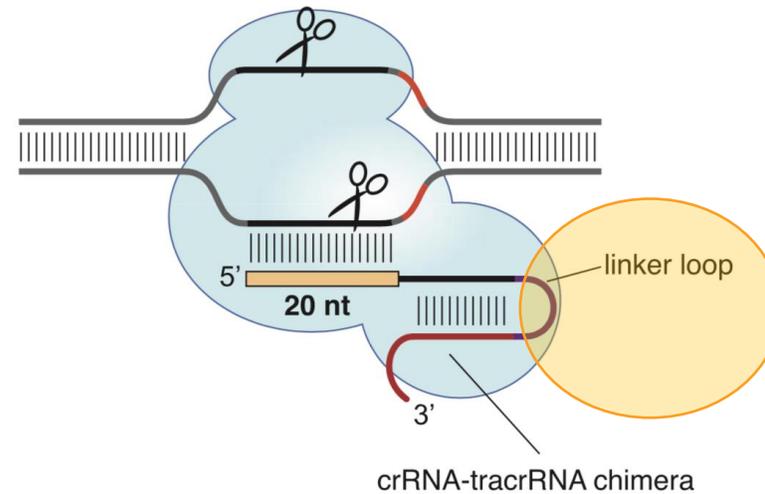


# Transforming bacterial CRISPR-Cas9 in a technology for genome editing

Cas9 programmed by crRNA:tracrRNA duplex



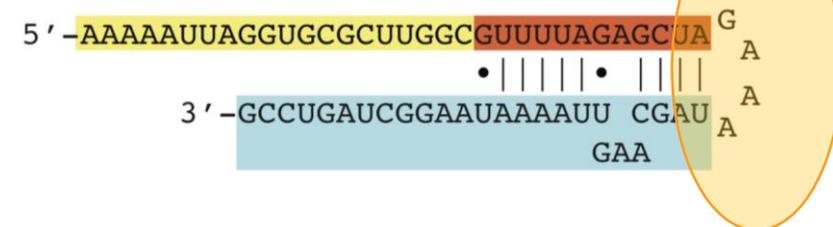
Cas9 programmed by single chimeric RNA



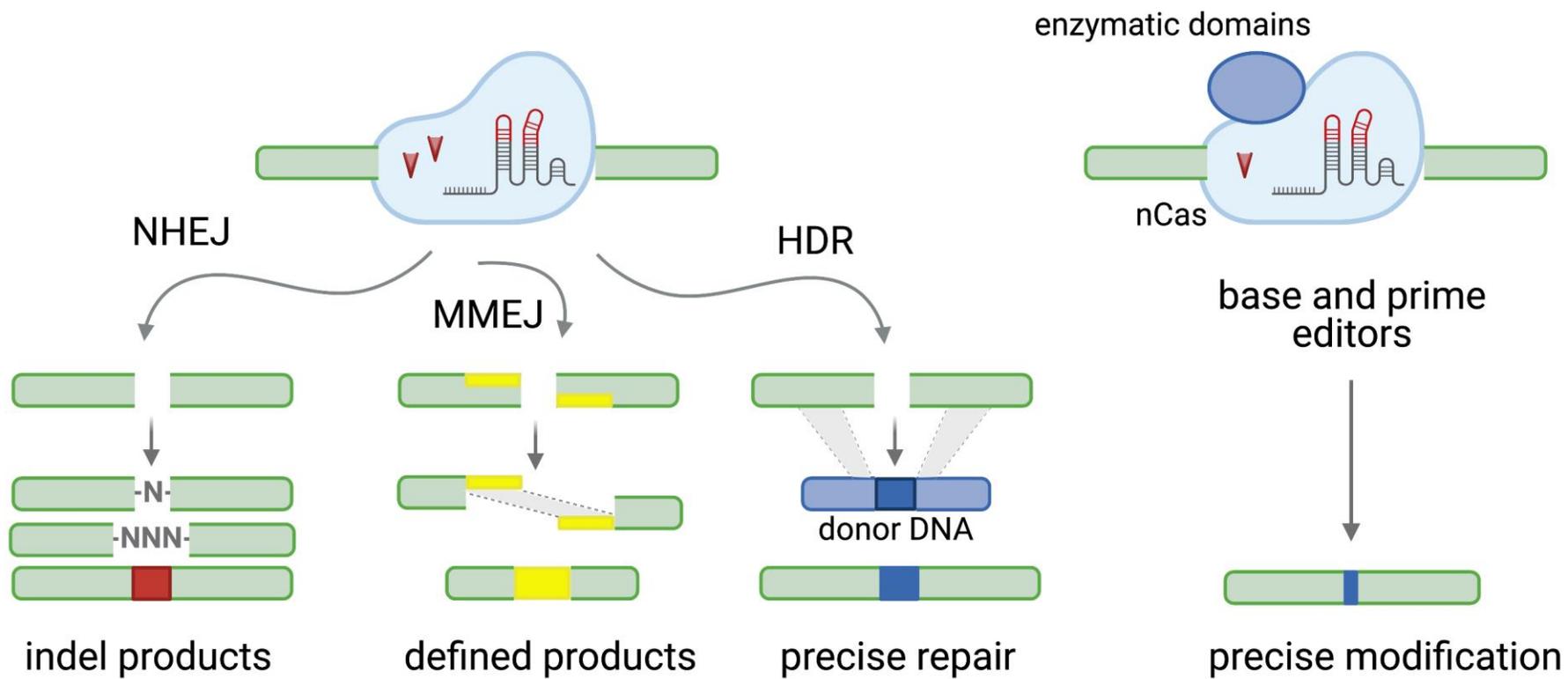
## A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity

Martin Jinek,<sup>1,2\*</sup> Krzysztof Chylinski,<sup>3,4\*</sup> Ines Fonfara,<sup>4</sup> Michael Hauer,<sup>2,†</sup> Jennifer A. Doudna,<sup>1,2,5,6,‡</sup> Emmanuelle Charpentier<sup>4,‡</sup>

### chimera A



# Editing outcome

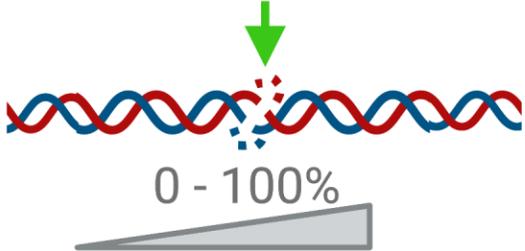


Le proprietà della tecnologia CRISPR-Cas

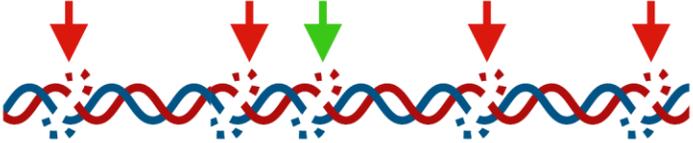
Considerazioni in-vitro

# Properties of CRISPR genome editing technology

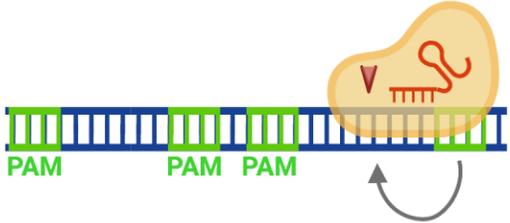
On target activity  
*efficiency?*



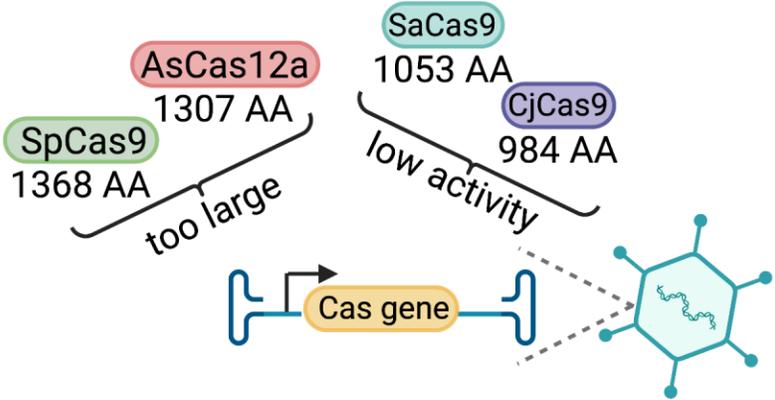
Off target activity  
*where else?*



Targeting range (PAM constraints)  
*where?*

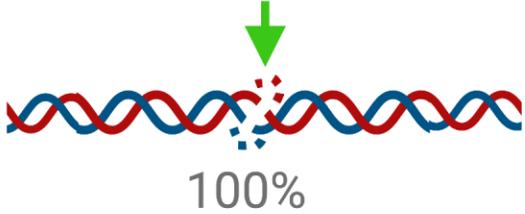


Delivery (Cas molecular weight)  
*how to deliver?*

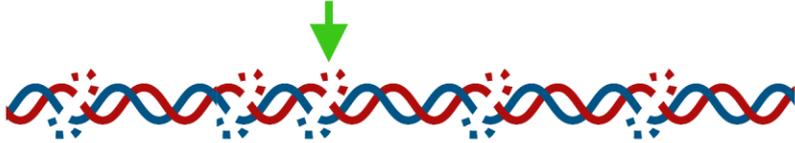


# Properties of CRISPR genome editing technology

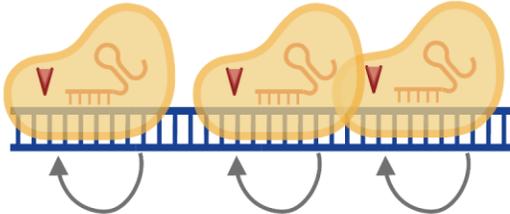
On target activity



Off target activity



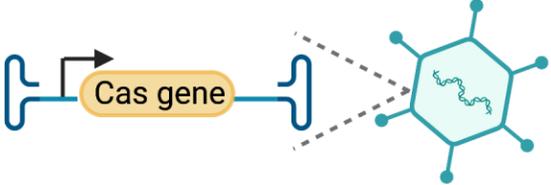
Targeting range (PAM constraints)

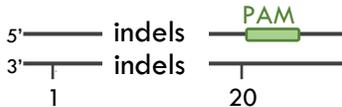
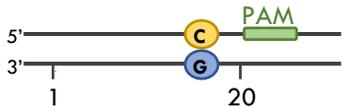
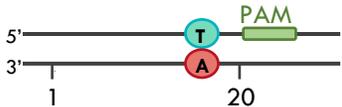
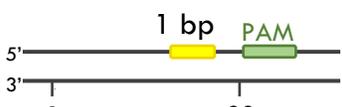
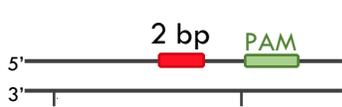
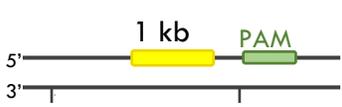
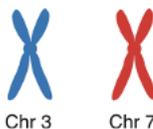
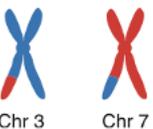


Delivery (Cas molecular weight)



small molecular weight

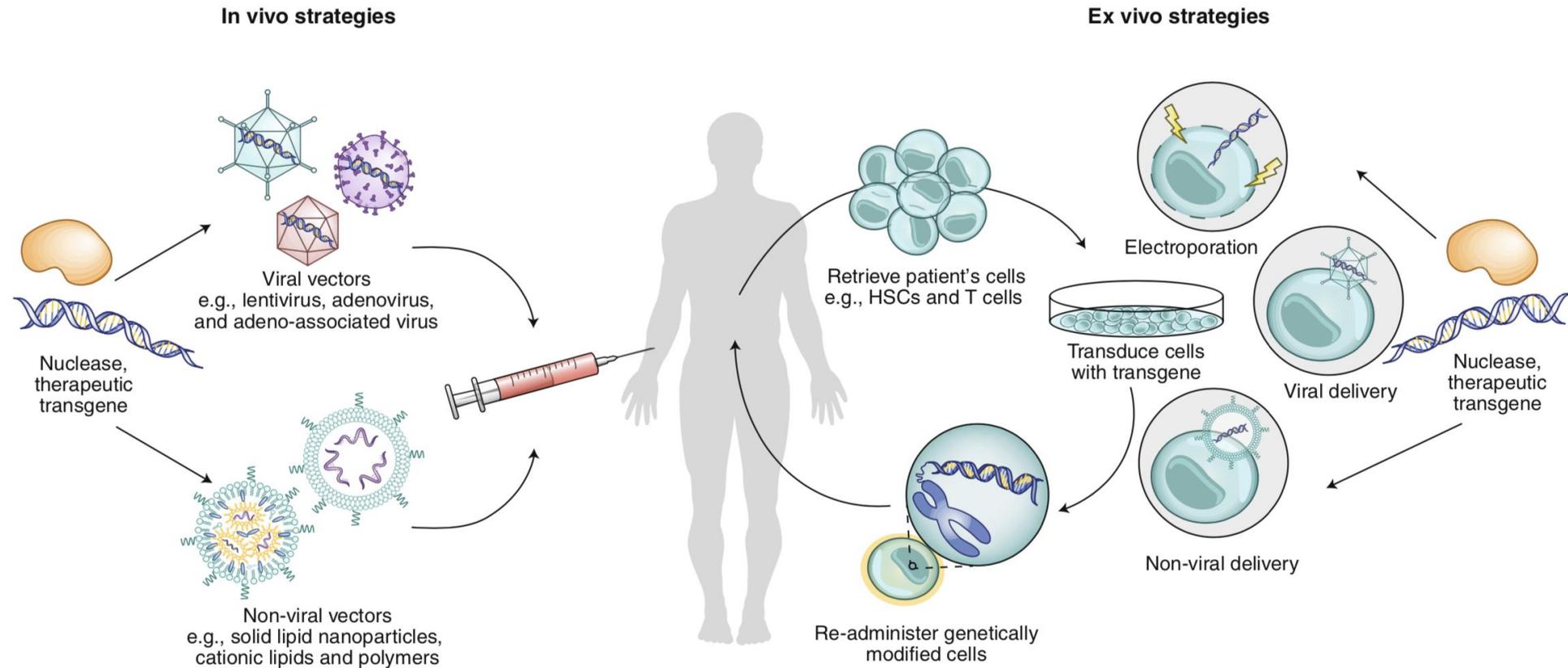


Edit types	Starting sequence	Desired product	Methods	Byproducts
<b>Stochastic indels</b>			Cas nuclease	Diverse indels not controllable
<b>PAM-distal transition point mutations</b>			Base editors	Bystander mutations
<b>PAM-proximal transition point mutations</b>			Cas nuclease HDR Prime editor	Extra indels
<b>Small insertions (e.g. 1-40 bp)</b>			Cas nuclease HDR Prime editor	Extra indels
<b>Small deletions (e.g. 1-80 bp)</b>			Cas nuclease HDR Prime editor	Extra indels
<b>Large insertions (&gt;30 bp)</b>			Cas nuclease HDR Prime editor EJ Cas transp/recomb	Extra indels, wrong insert orientation, multiple insertions, vector insertions
<b>Large deletions (&gt;40 bp)</b>			Cas nuclease EJ Cas nuclease HDR	Indels at cut sites, inverted sequence
<b>Cromosomal translocations</b>			Cas nuclease EJ Cas nuclease HDR	Indels at translocation junction, indels at sites without translocation

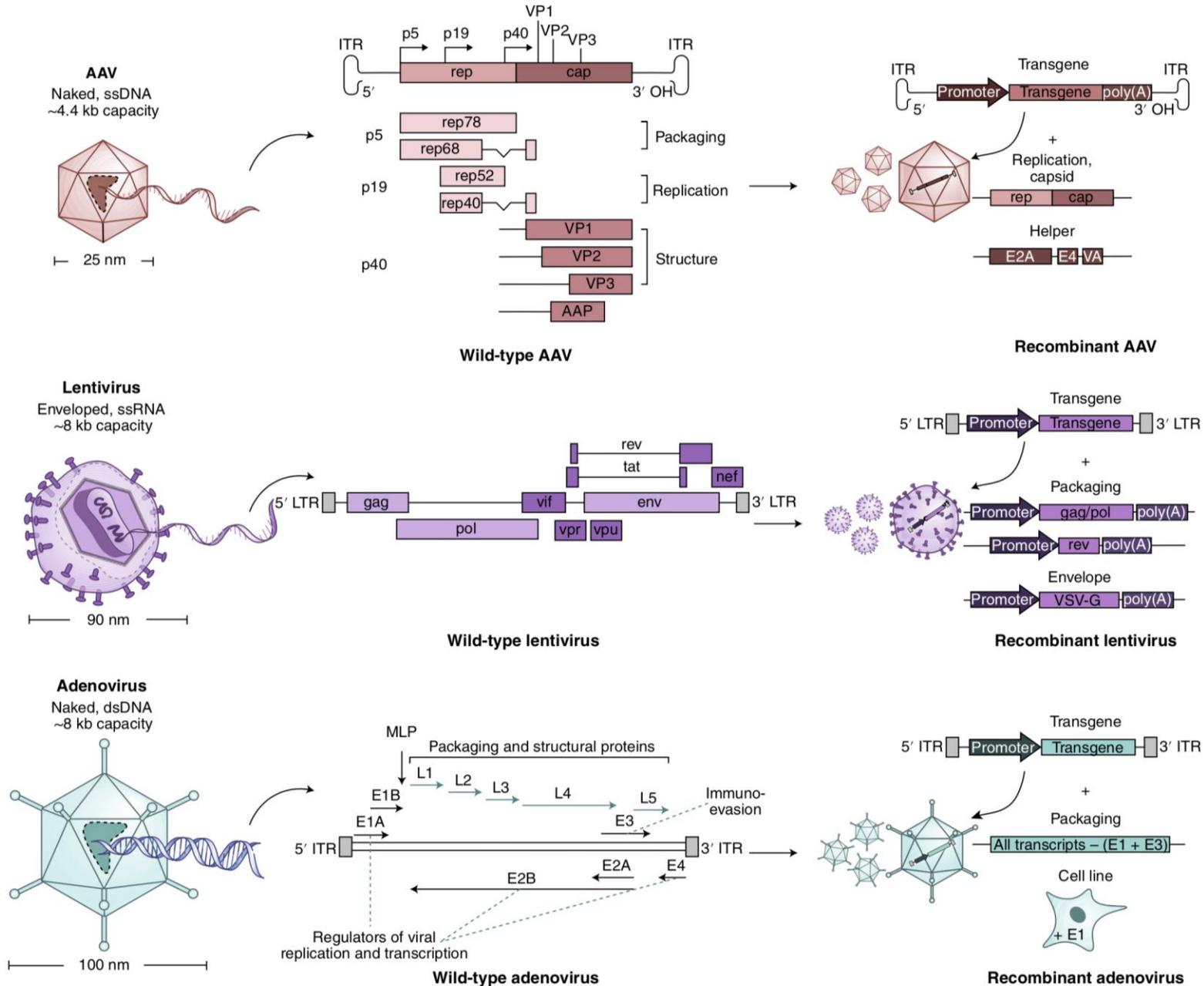
Le proprietà della tecnologia CRISPR-Cas

Considerazioni in-vivo

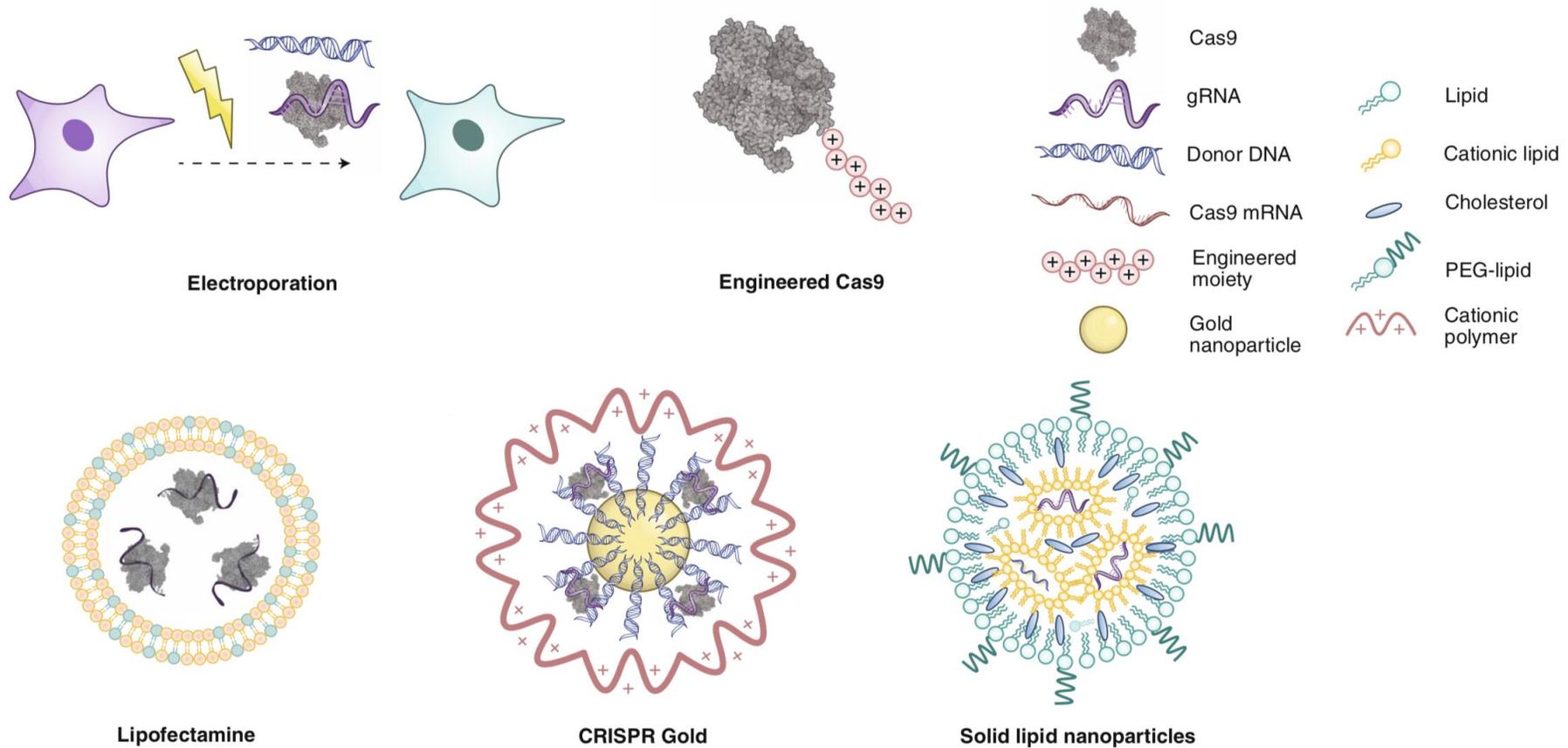
# Therapeutic genome editing strategies



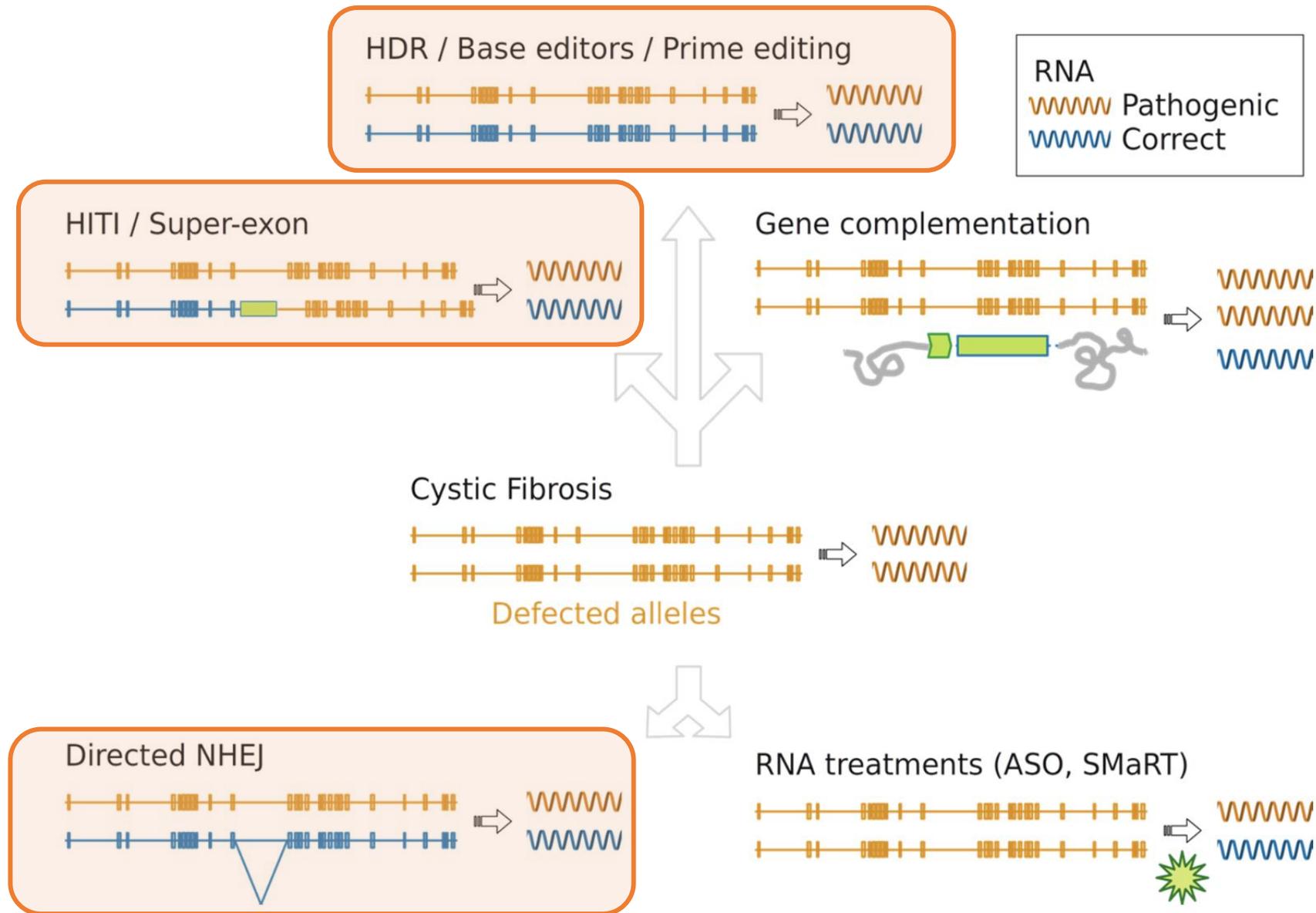
# Viral methods to deliver genome editing cargo



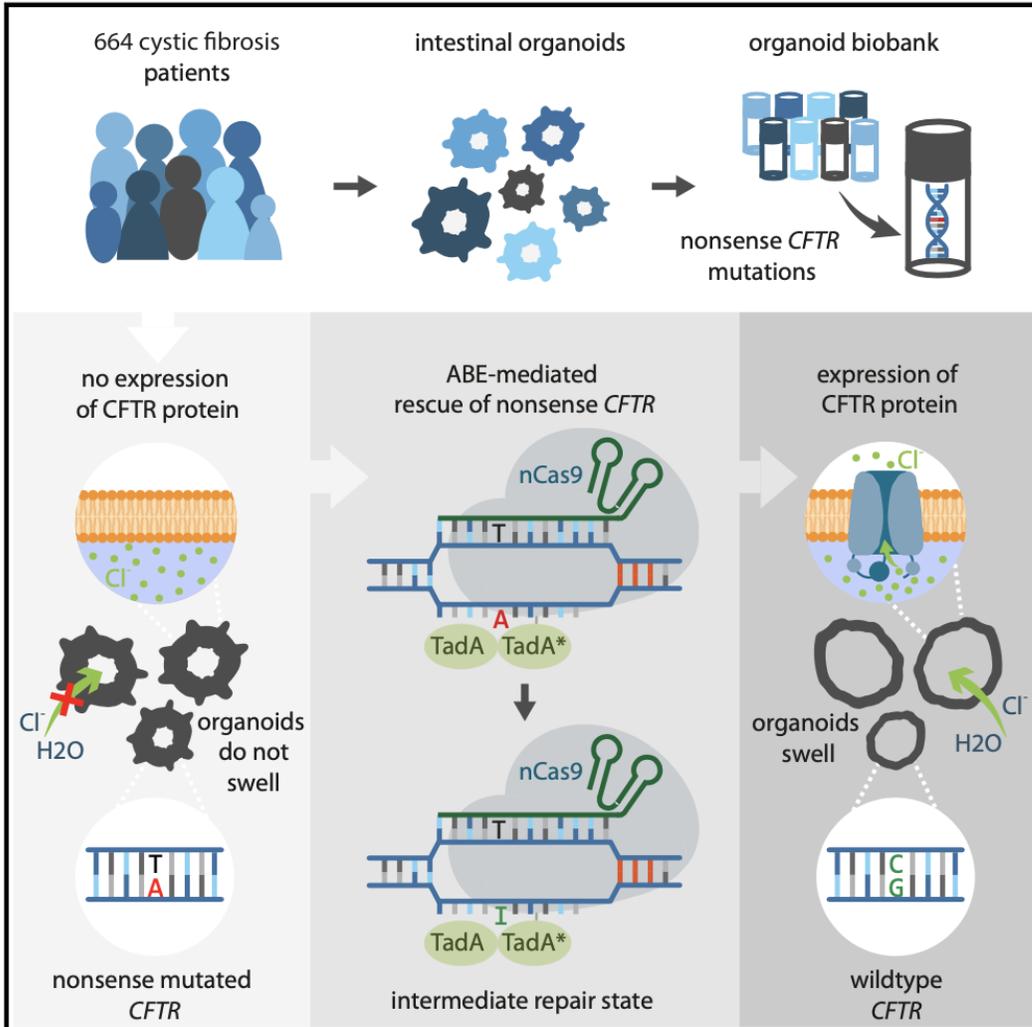
# Non-viral methods to deliver genome editing cargo



# Gene therapy for Cystic Fibrosis



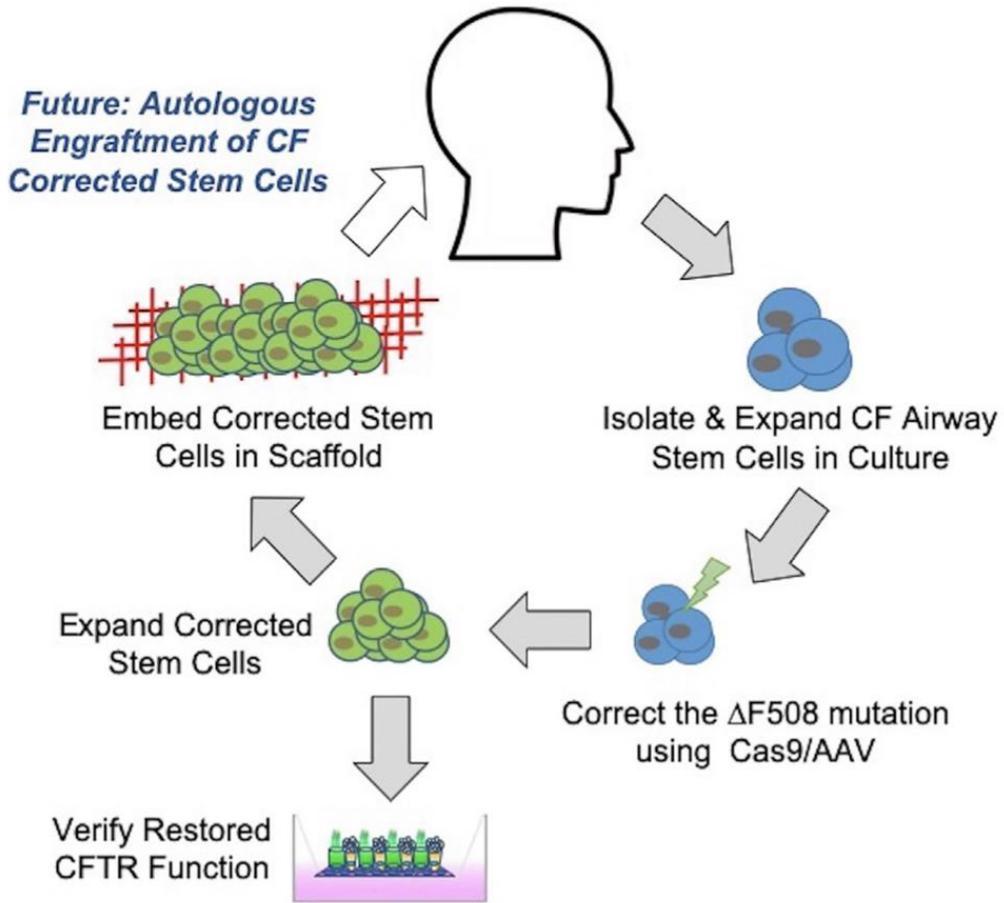
# Second generation CRISPR-Cas tools: Base-editors and prime editors



CRISPR-Based Adenine Editors Correct Nonsense Mutations in a Cystic Fibrosis Organoid Biobank- Cell Stem Cells, 2020-Hans Clevers

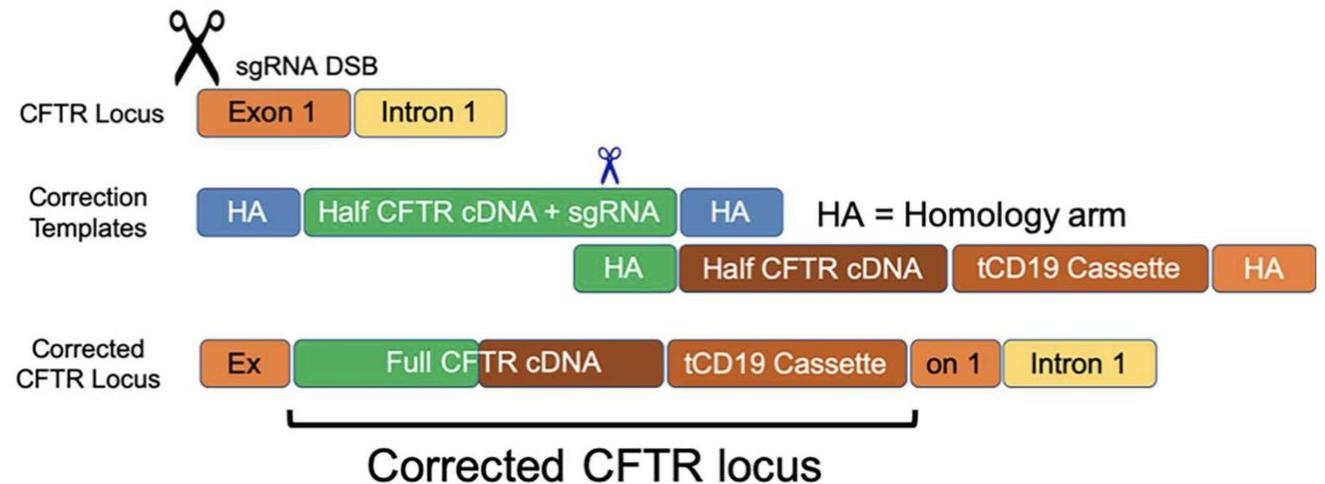
# Ex-vivo gene replacement through CRISPR-Cas induced homologous recombination

## Ex-Vivo Correction of Cystic Fibrosis (CF) Mutations in Airway Stem Cells



Targeted replacement of full-length CFTR in human airway stem cells by CRISPR/ Cas9 for pan-mutation correction in the endogenous locus – Mol Ther- 2021 M. Porteus

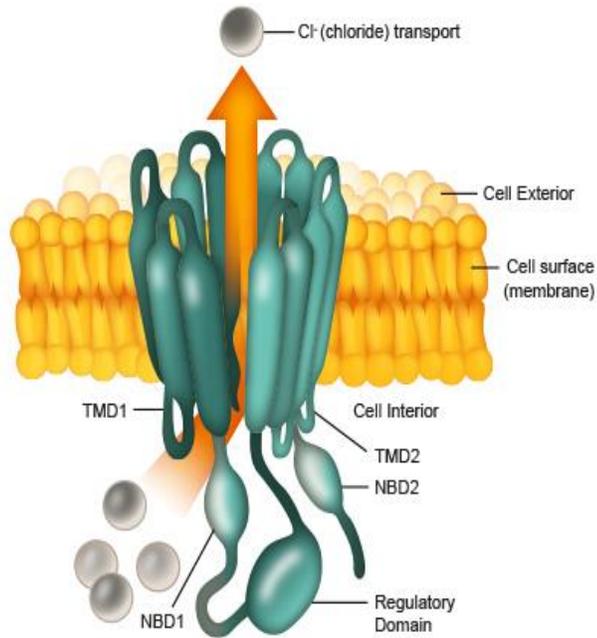
High-Efficiency, Selection-free Gene Repair in Airway Stem Cells from Cystic Fibrosis Patients Rescues CFTR Function in Differentiated Epithelia – Cell Stem Cell 2020 M Porteus



Correzione di una mutazione di splicing  
3272-26A>G in CFTR tramite genome  
editing con CRISPR-Cas

# Cystic fibrosis

- autosomal recessive monogenetic disease caused by mutations in the **CFTR** gene
- ABC transporter-class ion channel that conducts chloride ions across epithelial cell membranes



**Cystic  
Fibrosis  
Transmembrane conductance  
Regulator**

Class of mutation

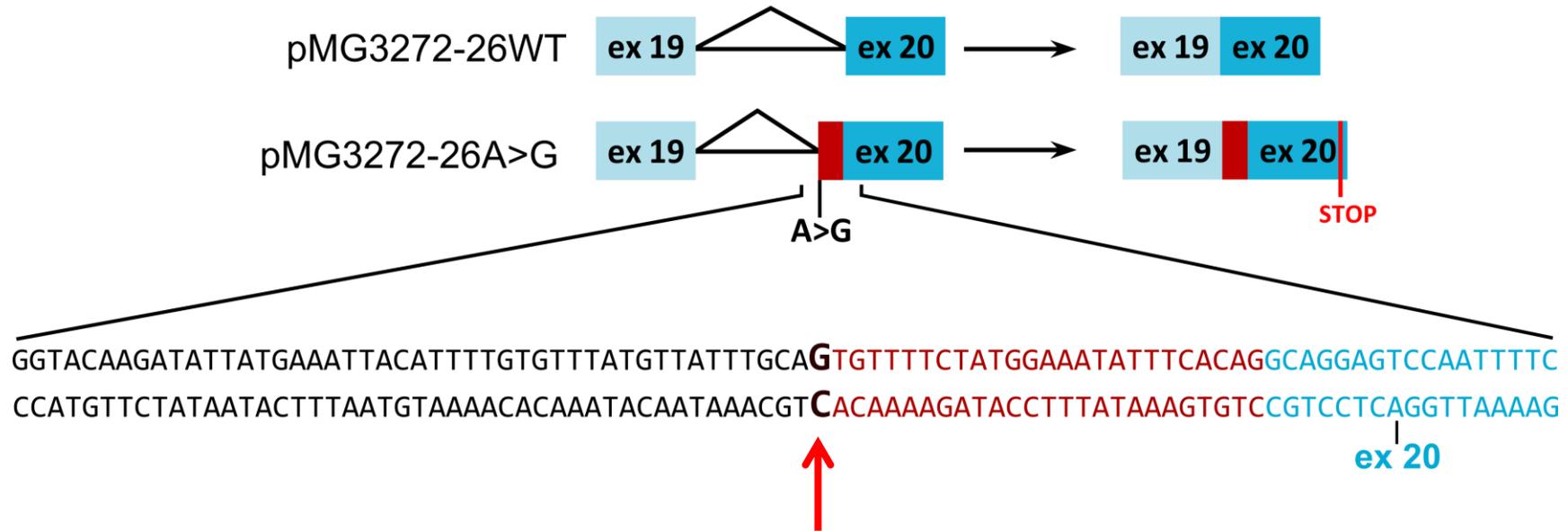
	Normal	I	II	III	IV	V	VI
Molecular defect	No synthesis	Block in processing	Block in regulation	Reduced conductance	Reduced synthesis	Reduced half-life	
Functional abnormality	Protein is not synthesized	Folding defect	Channel opening defect	Ion transport defect	Decreased protein synthesis	Decreased half-life of the protein	
Main mutations	Gly542X Trp128X Arg553X 621+1G→T	Phe508del Asn1303Lys Ile507del Arg560Thr	Gly551Asp Gly178Arg Gly551Ser Ser549Asn	Arg117His Arg347Pro Arg117Cys Arg334Trp	3849+10kbC→T 2789+5G→A 3272-26A →G 5T	4326delTC Gln1412X 4279insA	



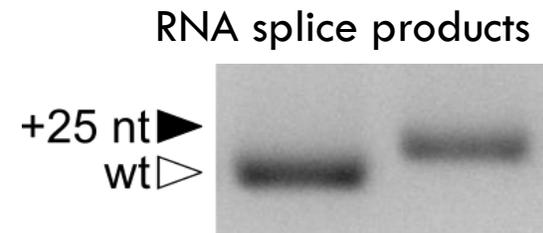
**Splicing defects**

# 3272-26A>G CF splicing mutation

## CFTR intron 19 MINIGENE models

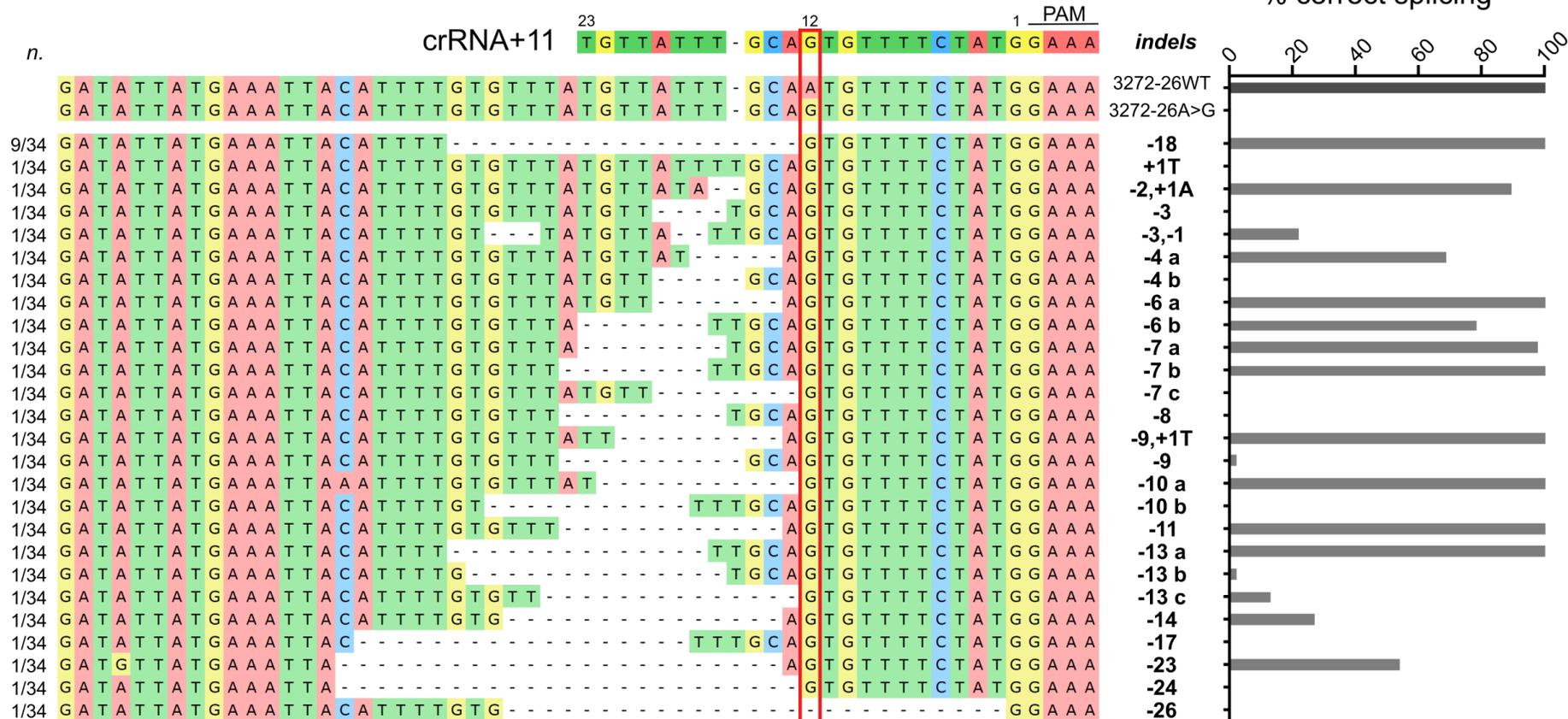


Alternative acceptor  
splice site

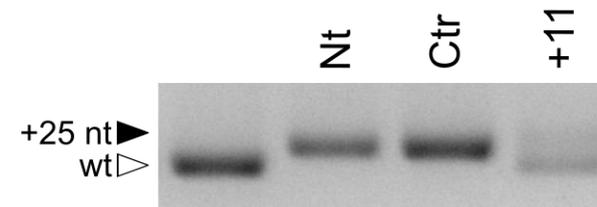


# AsCas12a-crRNA +11 editing

AsCas12a editing of *CFTR* pMG3272-26A>G



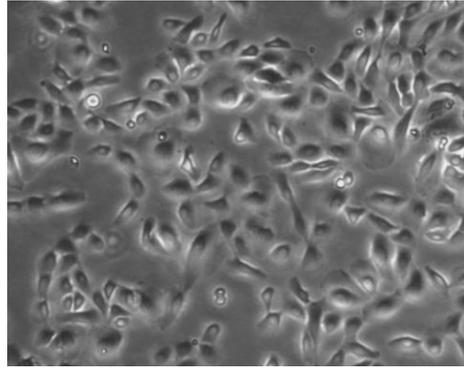
**68%** of analyzed editing events contributed to the effective restoration of *normal splicing*



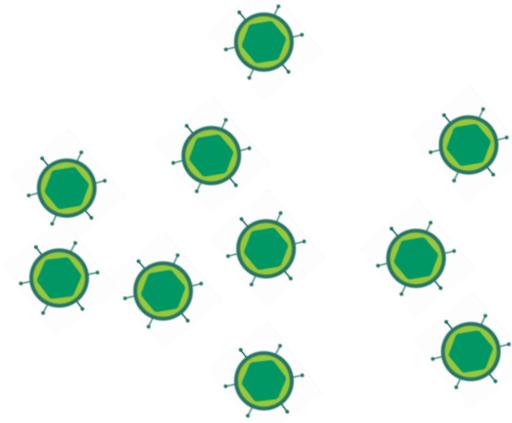
# CFTR splicing repair in primary airway cells



CF patient

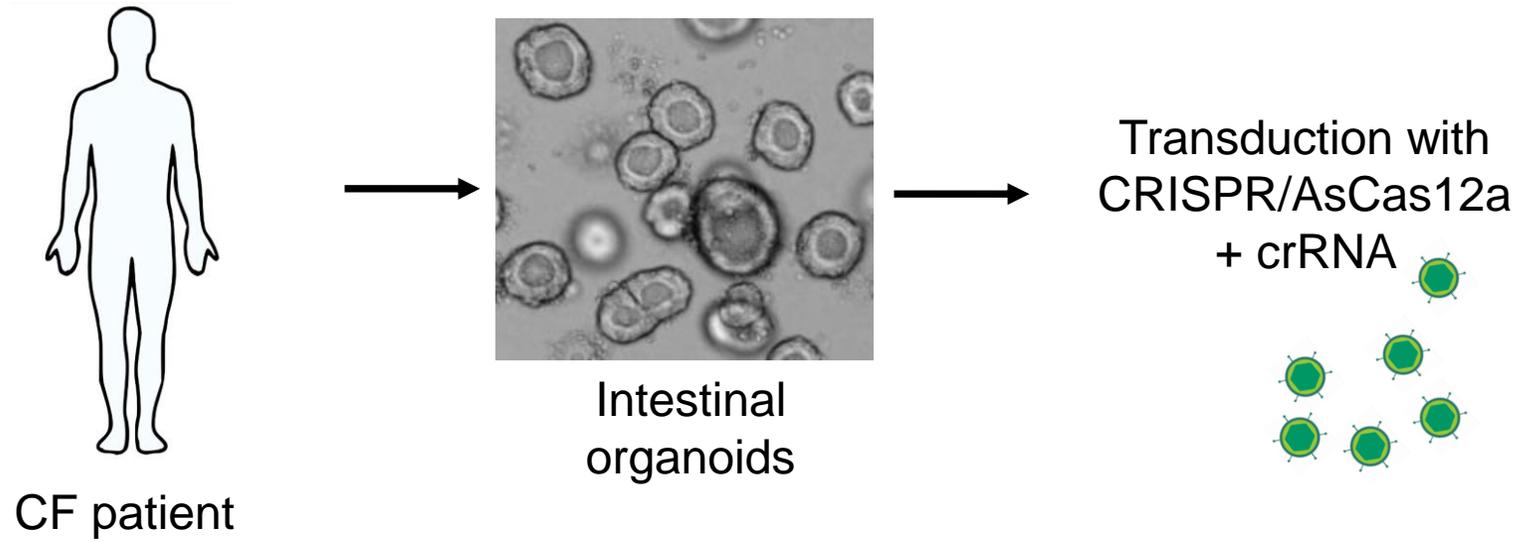


Primary  
airway cells

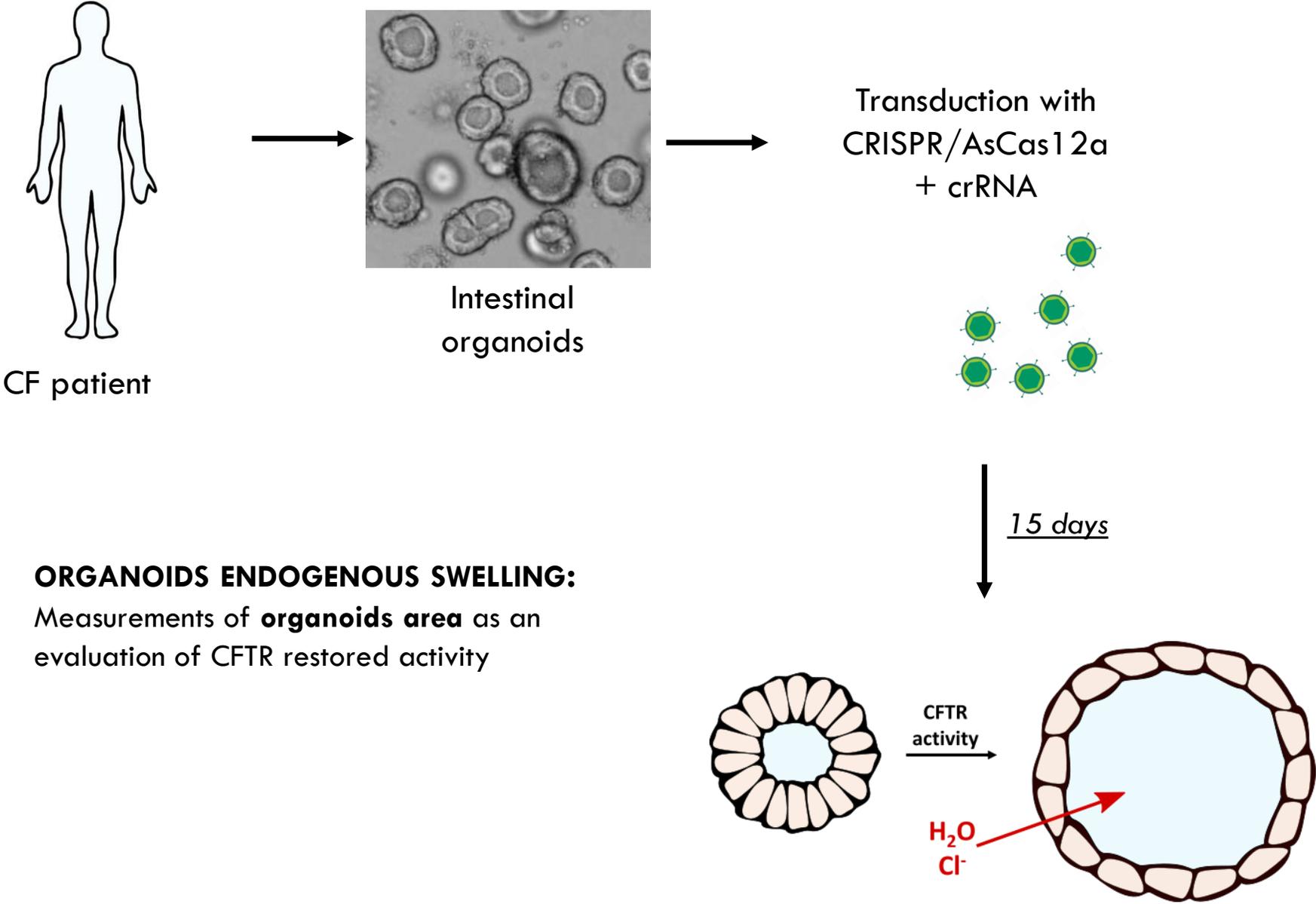


Lentiviral transduction with  
CRISPR/AsCas12a  
+ crRNA

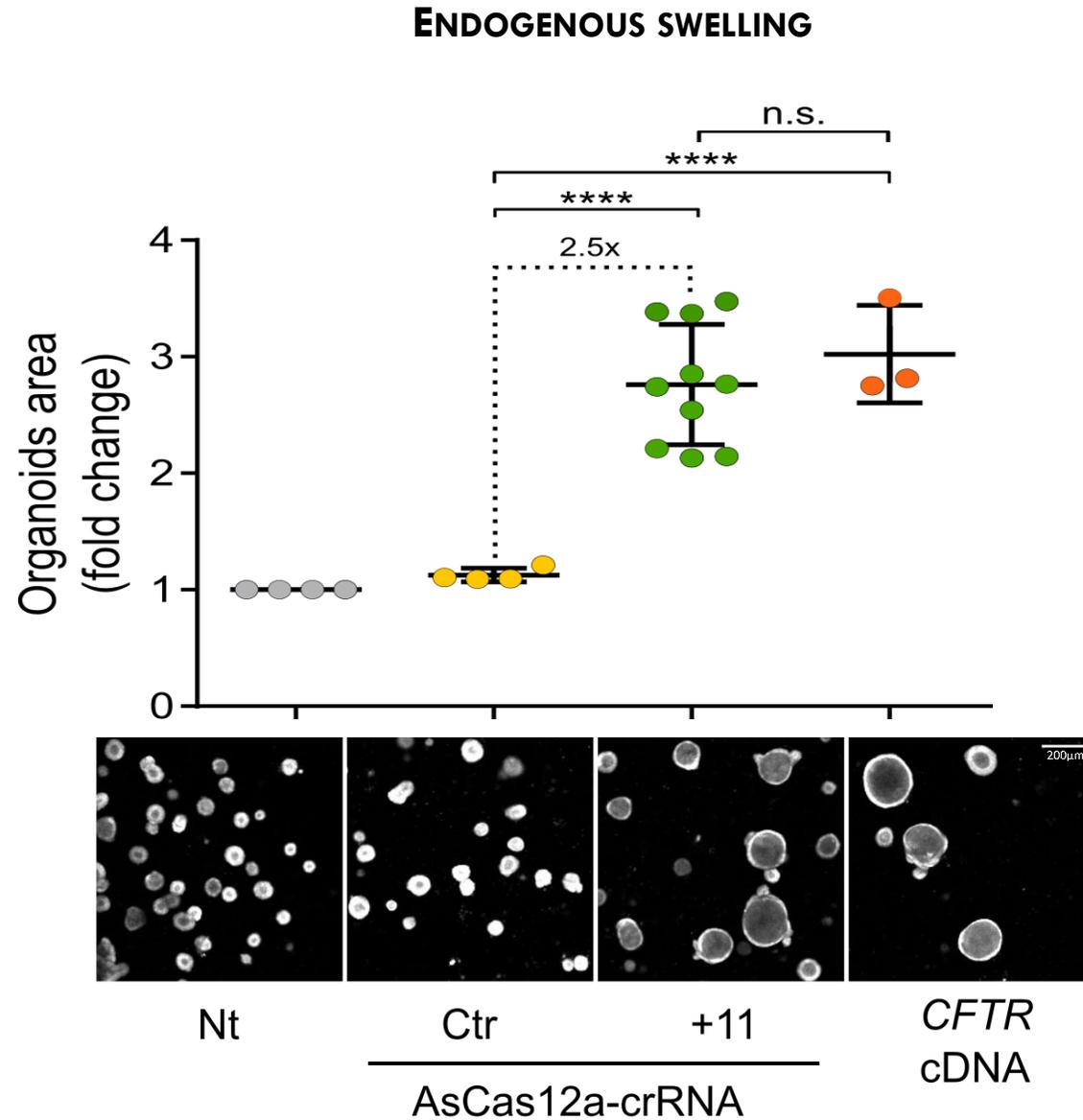
# ...and in intestinal organoids



# Organoids experiments



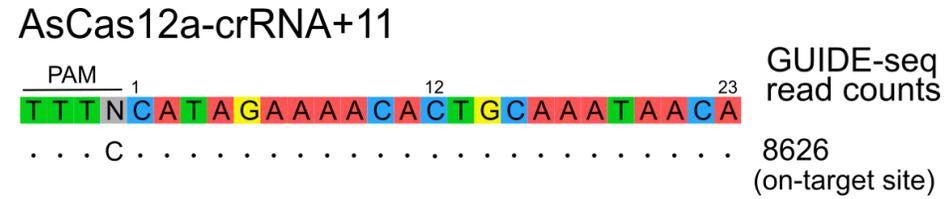
# AsCas12a editing in 3242-26A>G organoids



\*\*\*\* P<0,0001

# crRNA OFF target analysis

## GUIDE-SEQ METHOD

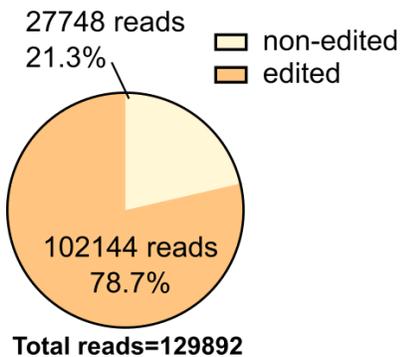


## TARGETED DEEP-SEQUENCING

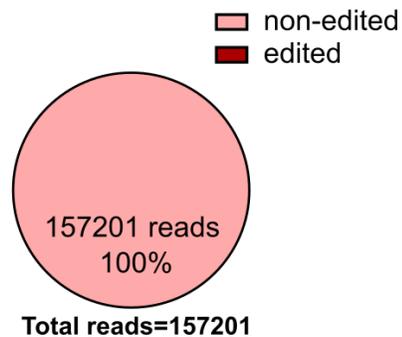
### Primary airway cells

### Organoids

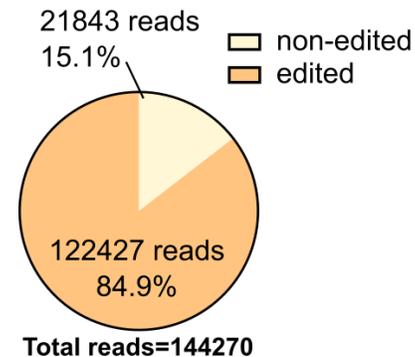
#### 3272-26A>G allele



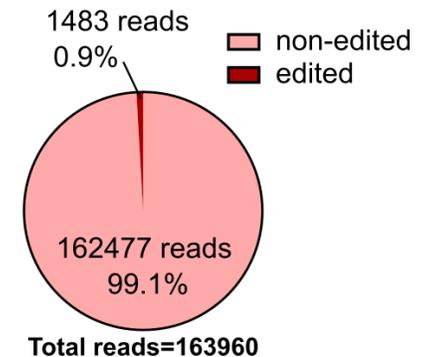
#### 3272-26WT allele



#### 3272-26A>G allele



#### 3272-26WT allele



## **Genome editing per fibrosi cistica. Dove siamo:**

La tecnologia CRISPR-Cas permette di correggere in-vitro la maggioranza (tutte) le mutazioni del gene *CFTR*.

Il successo in clinica della tecnologia dipende da:

**1) DELIVERY**

2) Precisione

3) Accessibilità del locus in cellule primarie (a seconda delle mutazioni)



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# Acknowledgements

## Molecular Virology Lab

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## Collaborations:

### CNR- Biophysics Institute

Daniele Arosio

### KU Leuven

Marianne Carlon

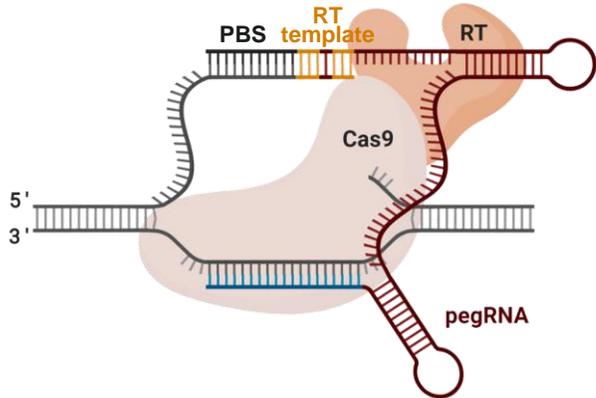
### Computational Metagenomics (UniTN)

Nicola Segata

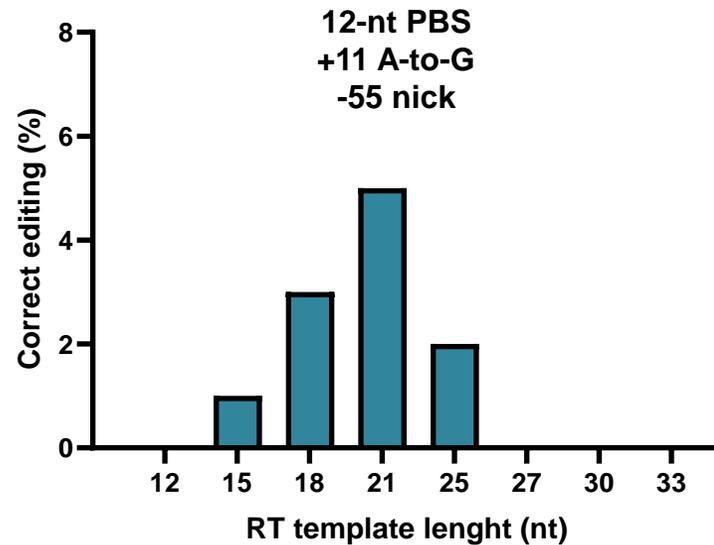




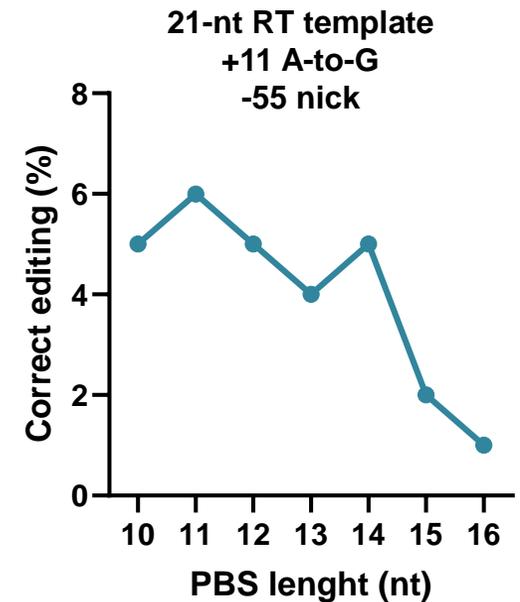
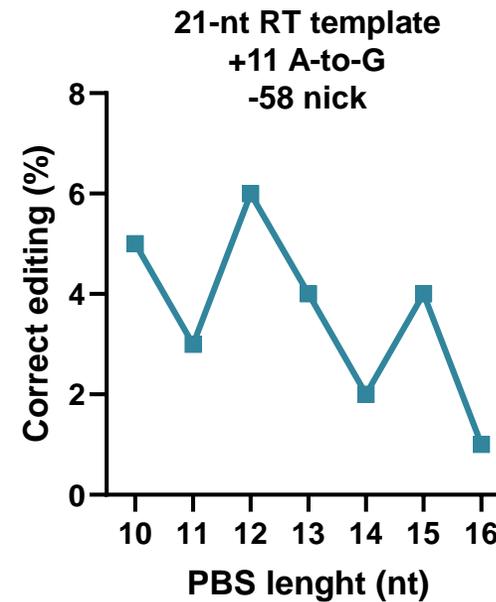
# Prime editing: 2789+5G>A: pegRNA optimization



RT template length



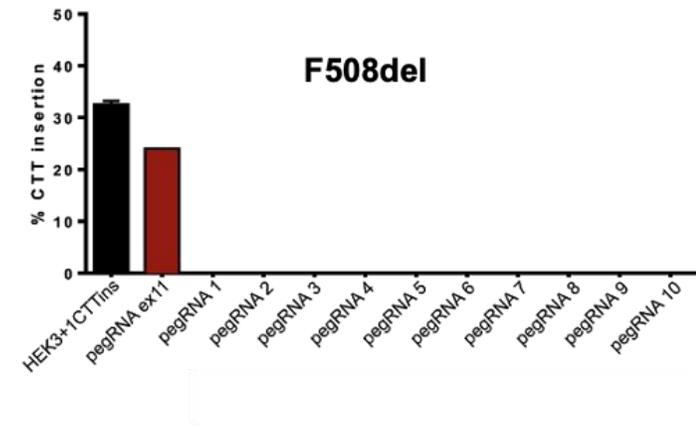
PBS length



A

pegRNAs	spacer sequence	3' extension	gRNA for PE3 strategy
HEK3 +1CTTins	GGCCAGACTGAGCACGTGA	TCTGCCATCA <b>AAG</b> CGTGCTCAGTCTG	GTCAACCAGTATCCCGGTGC
pegRNA ex11	GGGAGAACTGGAGCCTTCAG	TTACCCCTGAAG <b>AAG</b> GCTCCAGTTC	TGGAGATGTCCTCTTCTAGT
pegRNA1	TCTGTATCTATATTCATCAT	<b>TCTT</b> TGGTGTTTCCTATGATGAATATAG	CATTCTGTTCTCAGTTTTCC
pegRNA2	TCTGTATCTATATTCATCAT	<b>TCTT</b> TGGTGTTTCCTATGATGAATATAGAT	
pegRNA3	TCTGTATCTATATTCATCAT	<b>TCTT</b> TGGTGTTTCCTATGATGAATATAGATACA	
pegRNA4	TCTGTATCTATATTCATCAT	<b>TCTT</b> TGGTGTTTCCTATGATGAATATAGATACAGA	
pegRNA5	ACCATTAAAGAAAATATCAT	AGGAAACACCA <b>AAG</b> ATGATATTTCTT	TGGAGATGTCCTCTTCTAGT
pegRNA6	ACCATTAAAGAAAATATCAT	AGGAAACACCA <b>AAG</b> ATGATATTTCTTTA	
pegRNA7	ACCATTAAAGAAAATATCAT	AGGAAACACCA <b>AAG</b> ATGATATTTCTTTAAT	
pegRNA8	ACCATTAAAGAAAATATCAT	AGGAAACACCA <b>AAG</b> ATGATATTTCTTTAATGGT	
pegRNA9	CAGTTTTCTGGATTATGCC	ACCA <b>AAG</b> ATGATATTTCTTTAATGGTGCCAGGCATAATCCAGGAAAAC	TGGAGATGTCCTCTTCTAGT
pegRNA10	CAGTTTTCTGGATTATGCC	ACCA <b>AAG</b> ATGATATTTCTTTAATGGTGCCAGGCATAATCCAGGAAAAC	

B



# Use of CRISPR in biomedical field



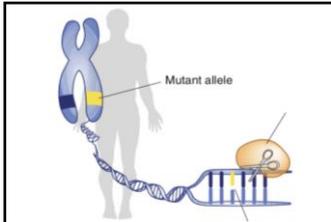
## Life science research

Genome editing for gene ko, screenings (drug), cell marking etc



## Animal model

Disease model to understand pathogenic mechanisms and therapeutic development.



## Gene therapy

Not just gene complementation but genes repair or ko of gain of function mutations



## Embryo genome editing

Treatments of genetic disease



## Virus detection and control

Identification and cleavage of viral nucleic acids or neutralization of viral co-factors (e.g. CCR5)

Genome editing  
TECHNOLOGY

Genome editing  
APPLICATIONS/CLINIC

**SLOW LANE**  
Next LTS in 3 months

**FAST LANE**  
Latest Release available



# Use of CRISPR



## **De-extinction**

Woolly mammoths (*Mammuthus primigenius*) to extinction.



## **Disease control**

Disease resistance is one of the most popular applications for CRISPR help to stem the dramatic loss of honeybees.



## **Better food production**

The US Food and Drug Administration approved the first transgenic animals for human consumption: fast-growing salmon.



## **Making drugs**

chickens with components required for CRISPR integrated directly into their genomes (CRISPR chickens) to edit chicken DNA: 'farmaceuticals' — drugs created using domesticated animals.



## **Vector control**

genetically modifying mosquitos to prevent the spread of diseases such as dengue or malaria.

## Take-home message

### Genome editing for cystic fibrosis:

Technological opportunities through CRISPR-Cas:  
**the majority/all mutations can be corrected this technology**

More advancements needed:

**1) DELIVERY**

2) Precision

3) Genomic locus accessibility

# Genome-editing in the literature

