

Il futuro del genome editing

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UNIVERSITY
OF TRENTO - Italy
Department CIBIO



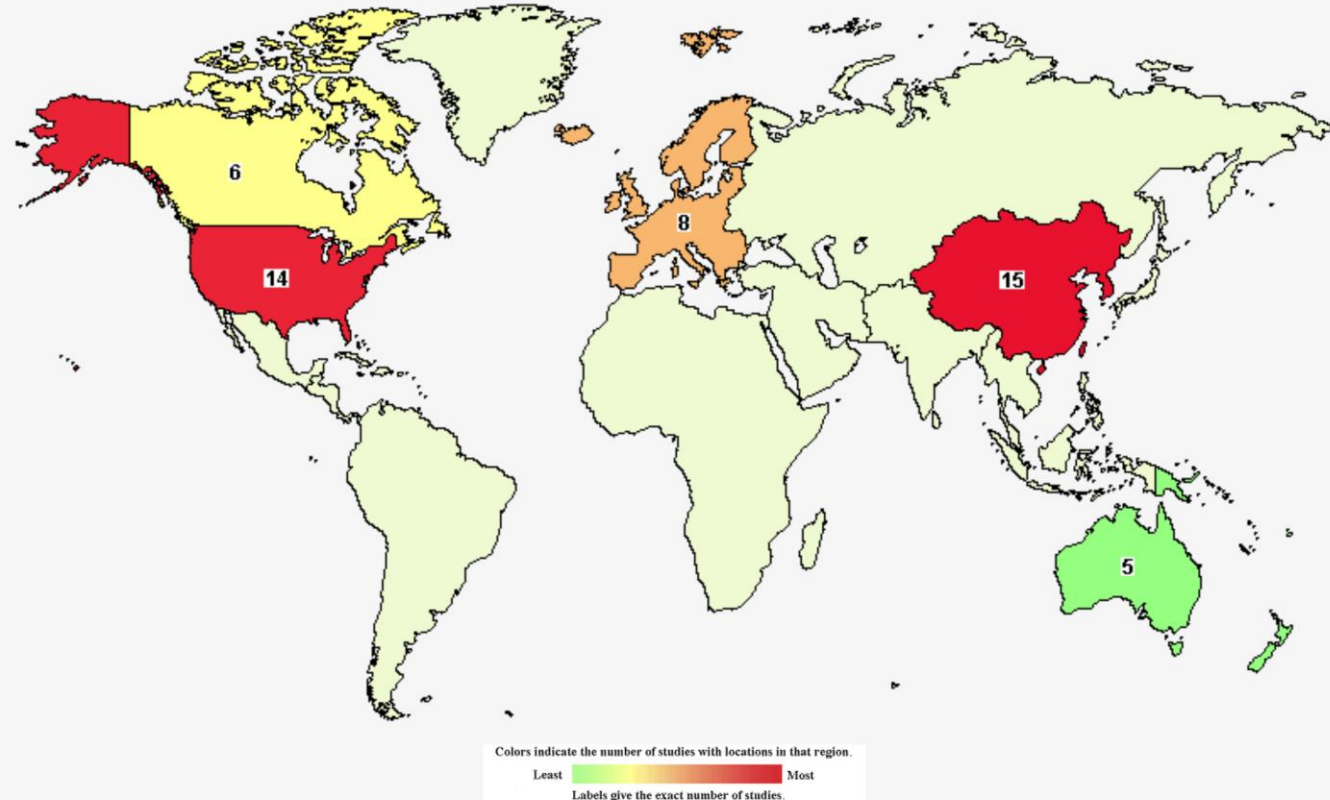
World map clinical trials using CRISPR-Cas9 technology



[List](#) [By Topic](#) [On Map](#) [Search Details](#)

[A similar map is available for all studies in ClinicalTrials.gov](#)

Click on the map below to show a more detailed map (when available) or search for studies (when map not available).



Source: <https://ClinicalTrials.gov>

Selected trials from the world list:

Europe:

- Beta-Thalassemia
- Sickle Cell Disease

US:

- Edited T cells (TCR and PD1 ko)
- Multiple Myeloma
- Melanoma
- Synovial Sarcoma
- Myxoid/Round Cell Liposarcoma

- Blindness (Leber Congenital Amaurosis)

China

- CAR-T cells (CD19, CD20, CD22)
- B Cell Leukemia
- B Cell Lymphoma

- CCR5 ko HIV-1-infection

- PD1 ko for EBV (Epstein Barr) malignancies (Gastric, Nasopharyngeal, Lymphoma)

October 2021: 37 studies

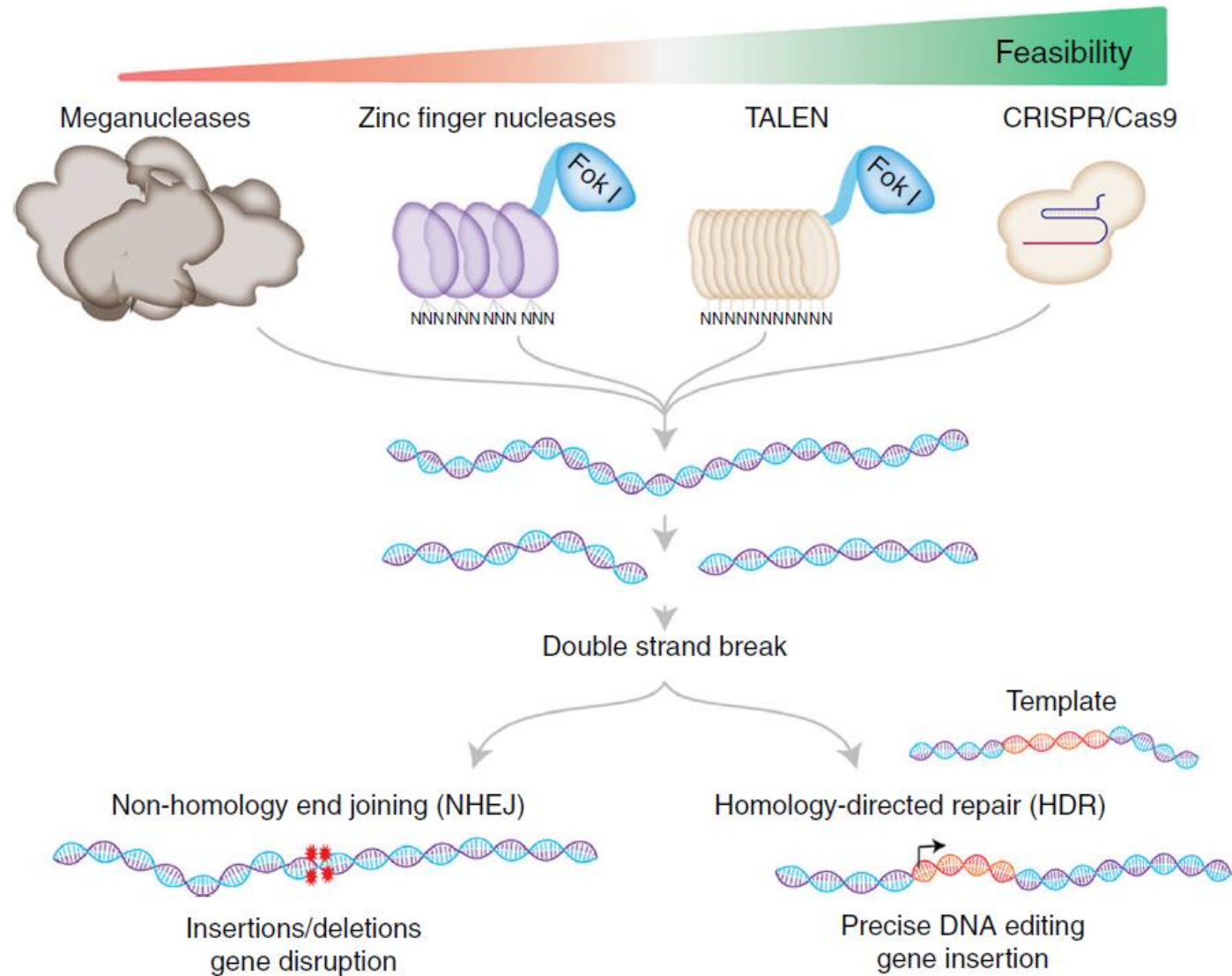
I. Background

- Avanzamenti e ostacoli del genome editing
- Proprietà in-vitro/in-vivo del genome editing
- Aggiornamenti di genome editing per la fibrosi cistica

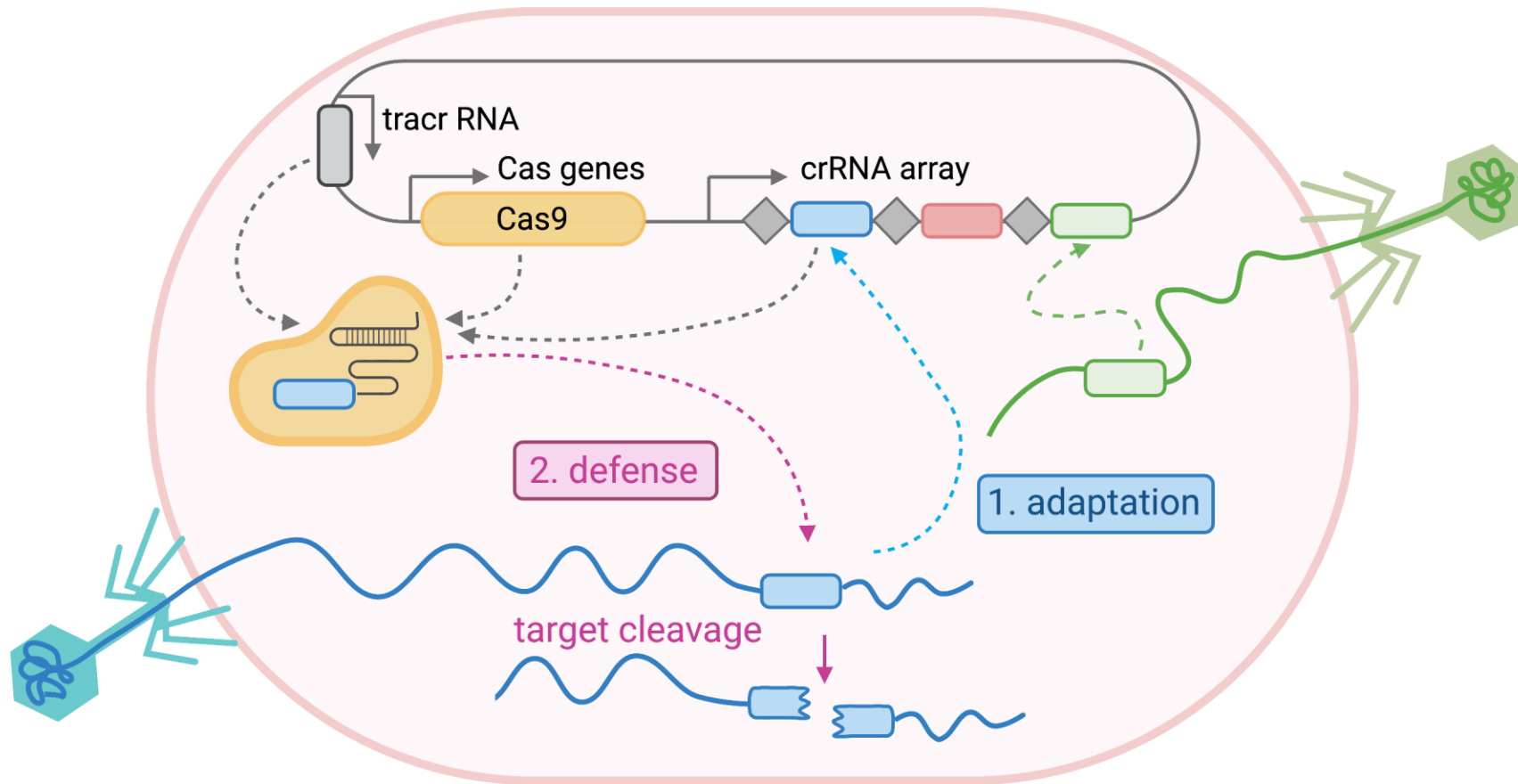
II. Esempio di correzione di mutazione di splicing

- Correzione della mutazione di splicing 3272-26A>G utilizzando CRISPR-Cas

Genome editing technology

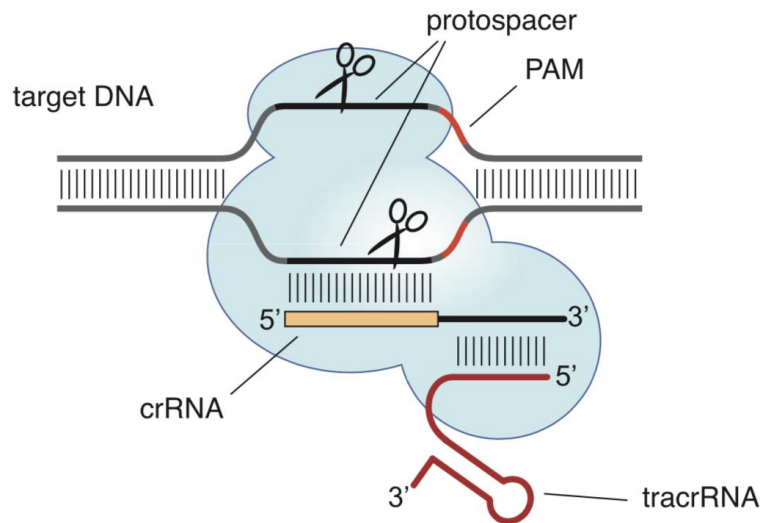


Host defense in bacteria

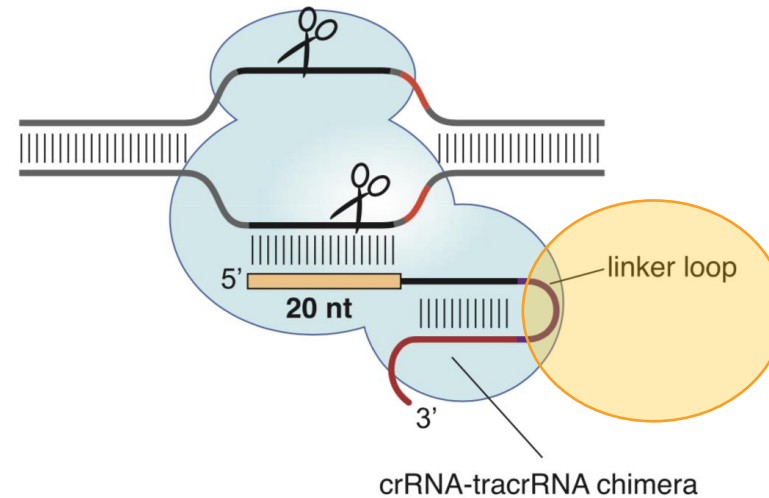


Transforming bacterial CRISPR-Cas9 in a technology for genome editing

Cas9 programmed by crRNA:tracrRNA duplex



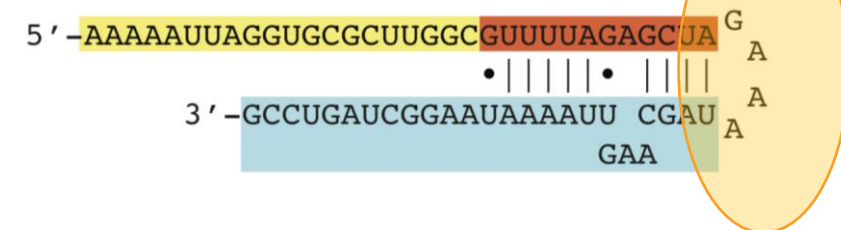
Cas9 programmed by single chimeric RNA



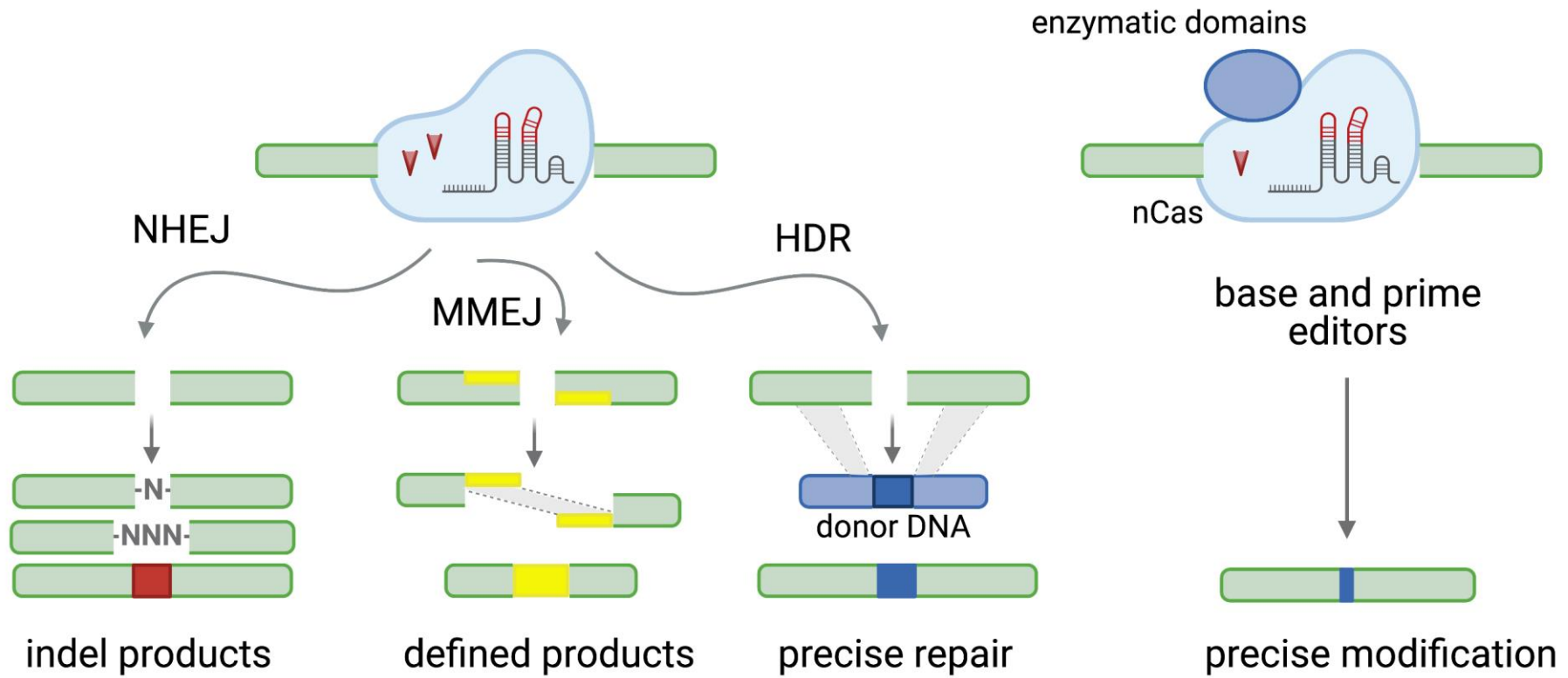
A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity

Martin Jinek,^{1,2*} Krzysztof Chylinski,^{3,4*} Ines Fonfara,⁴ Michael Hauer,^{2,†} Jennifer A. Doudna,^{1,2,5,6,‡} Emmanuelle Charpentier^{4,‡}

chimera A



Editing outcome

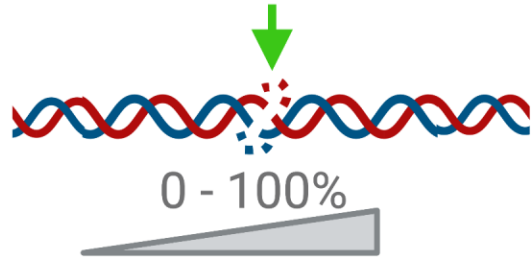


Le proprietà della tecnologia CRISPR-Cas

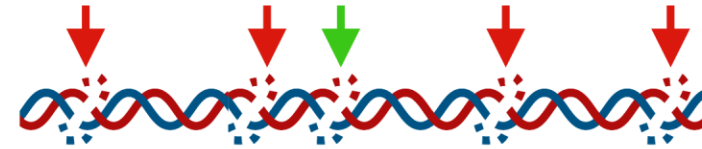
Considerazioni in-vitro

Properties of CRISPR genome editing technology

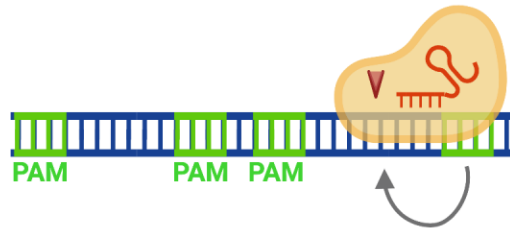
On target activity
efficiency?



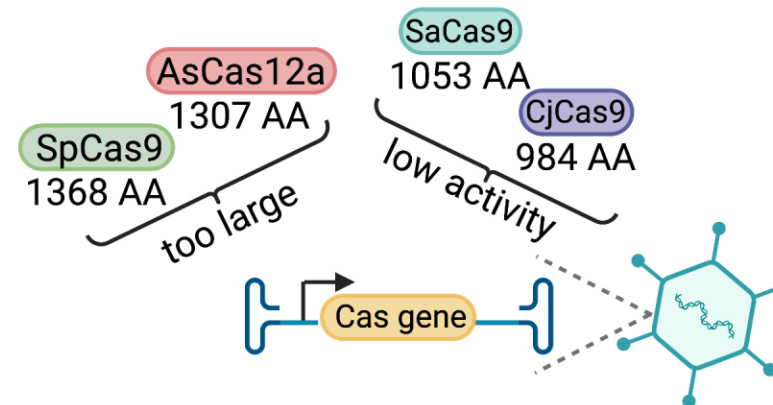
Off target activity
where else?



Targeting range (PAM constraints)
where?

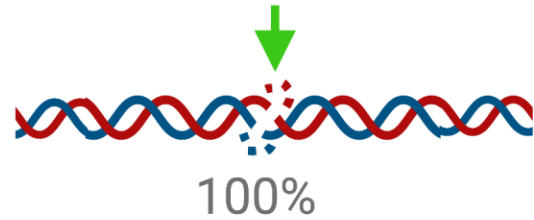


Delivery (Cas molecular weight)
how to deliver?

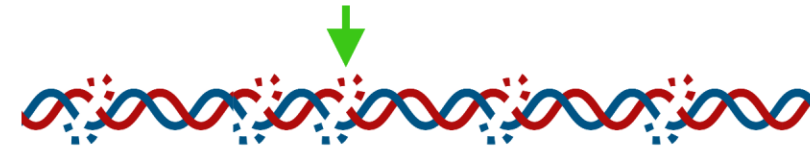


Properties of CRISPR genome editing technology

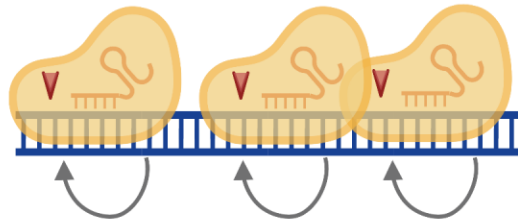
On target activity



Off target activity



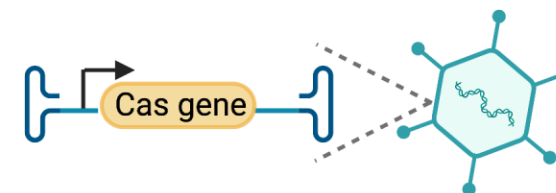
Targeting range (PAM constraints)



Delivery (Cas molecular weight)



small molecular weight

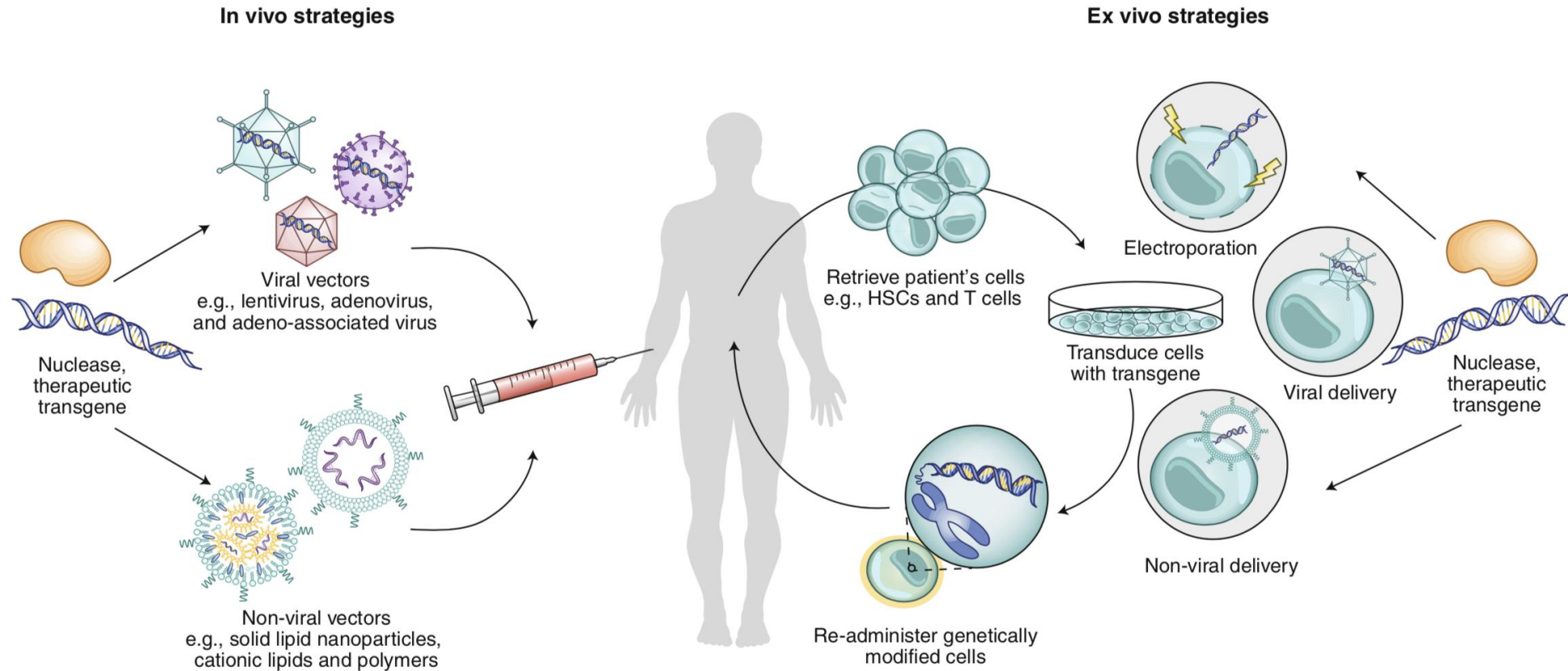


Edit types	Starting sequence	Desired product	Methods	Byproducts
Stochastic indels			Cas nuclease	Diverse indels not controllable
PAM-distal transition point mutations			Base editors	Bystander mutations
PAM-proximal transition point mutations			Cas nuclease HDR Prime editor	Extra indels
Small insertions (e.g. 1-40 bp)			Cas nuclease HDR Prime editor	Extra indels
Small deletions (e.g. 1-80 bp)			Cas nuclease HDR Prime editor	Extra indels
Large insertions (>30 bp)			Cas nuclease HDR Prime editor EJ Cas transp/recomb	Extra indels, wrong insert orientation, multiple insertions, vector insertions
Large deletions (>40 bp)			Cas nuclease EJ Cas nuclease HDR	Indels at cut sites, inverted sequence
Cromosomal translocations			Cas nuclease EJ Cas nuclease HDR	Indels at translocation junction, indels at sites without translocation

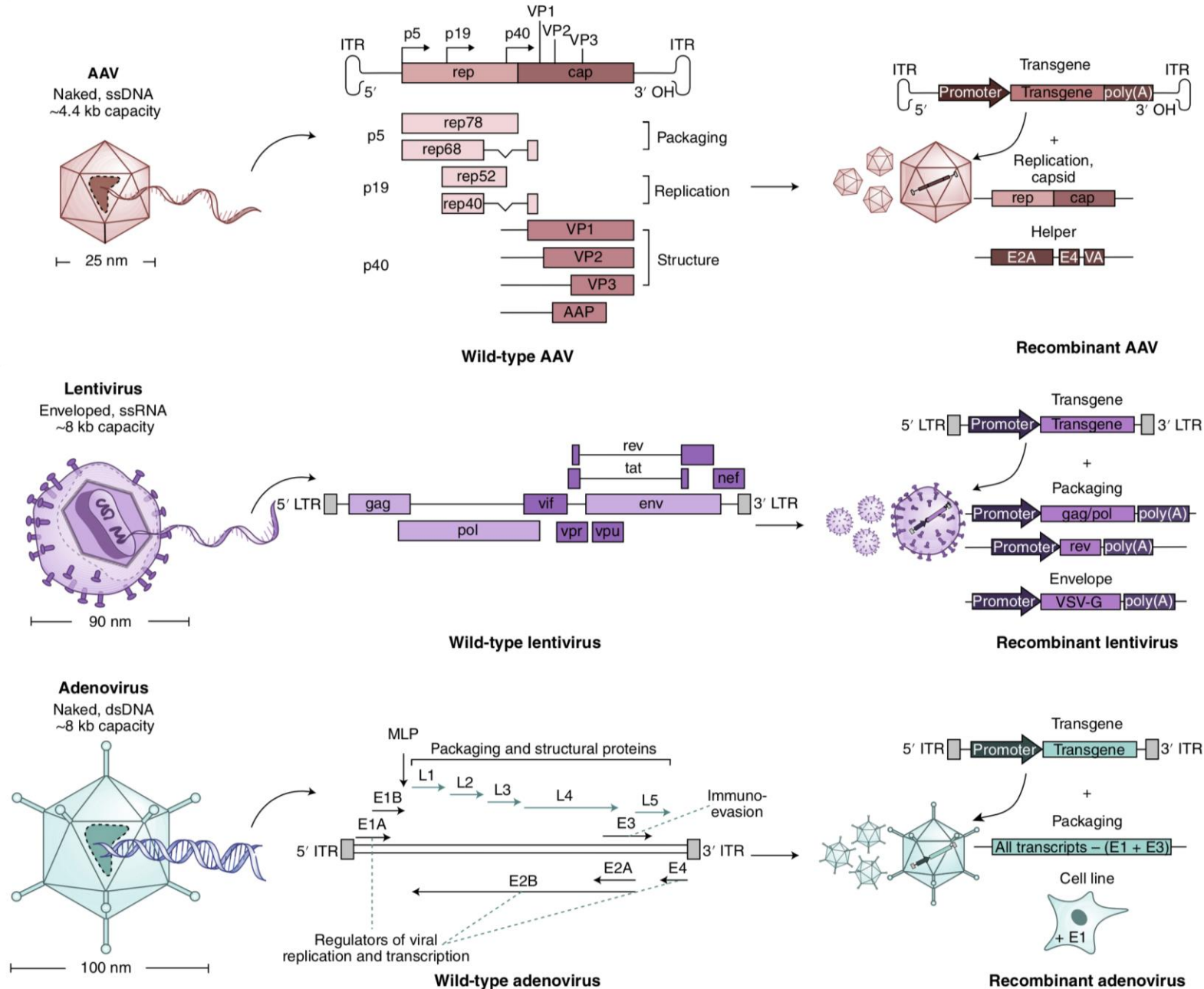
Le proprietà della tecnologia CRISPR-Cas

Considerazioni in-vivo

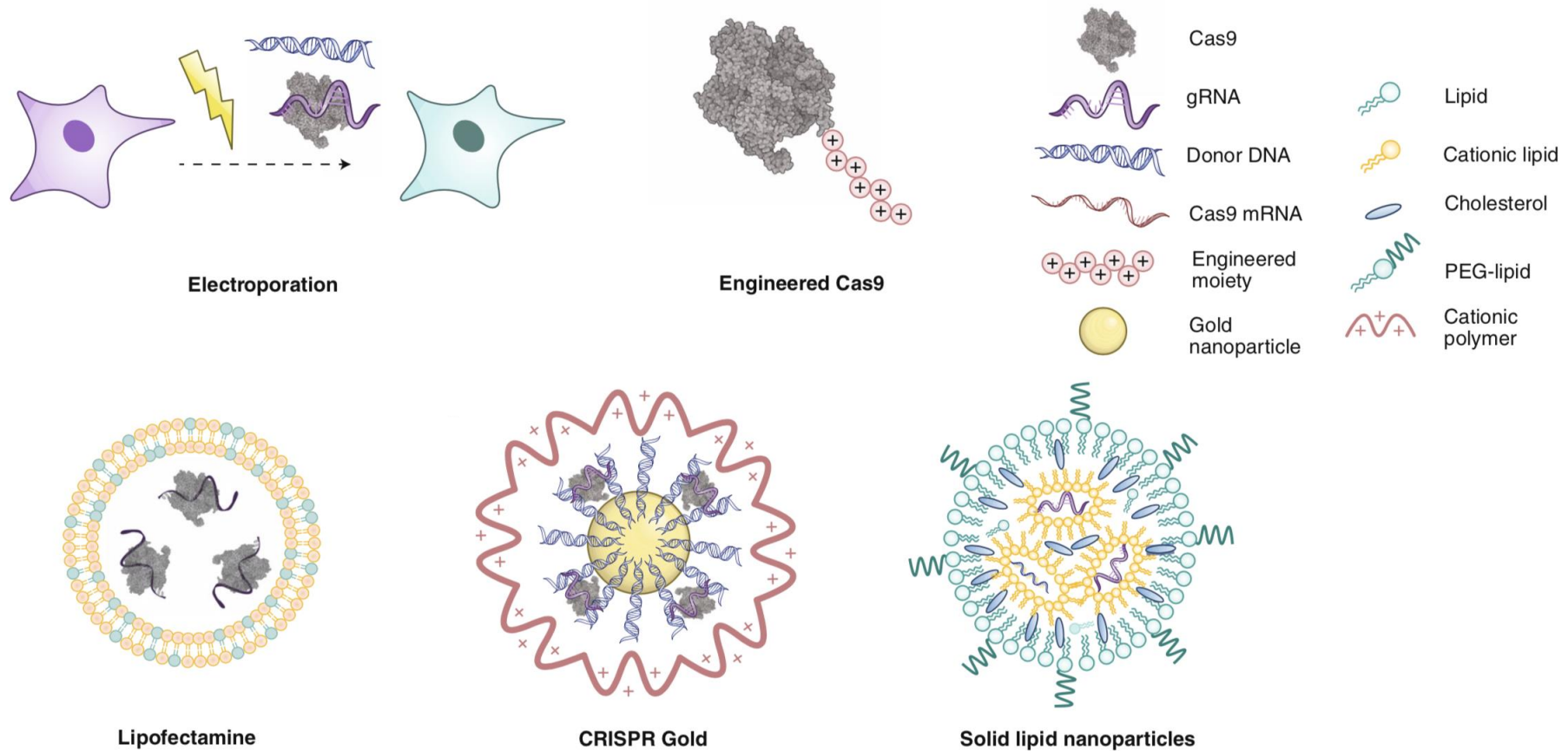
Therapeutic genome editing strategies



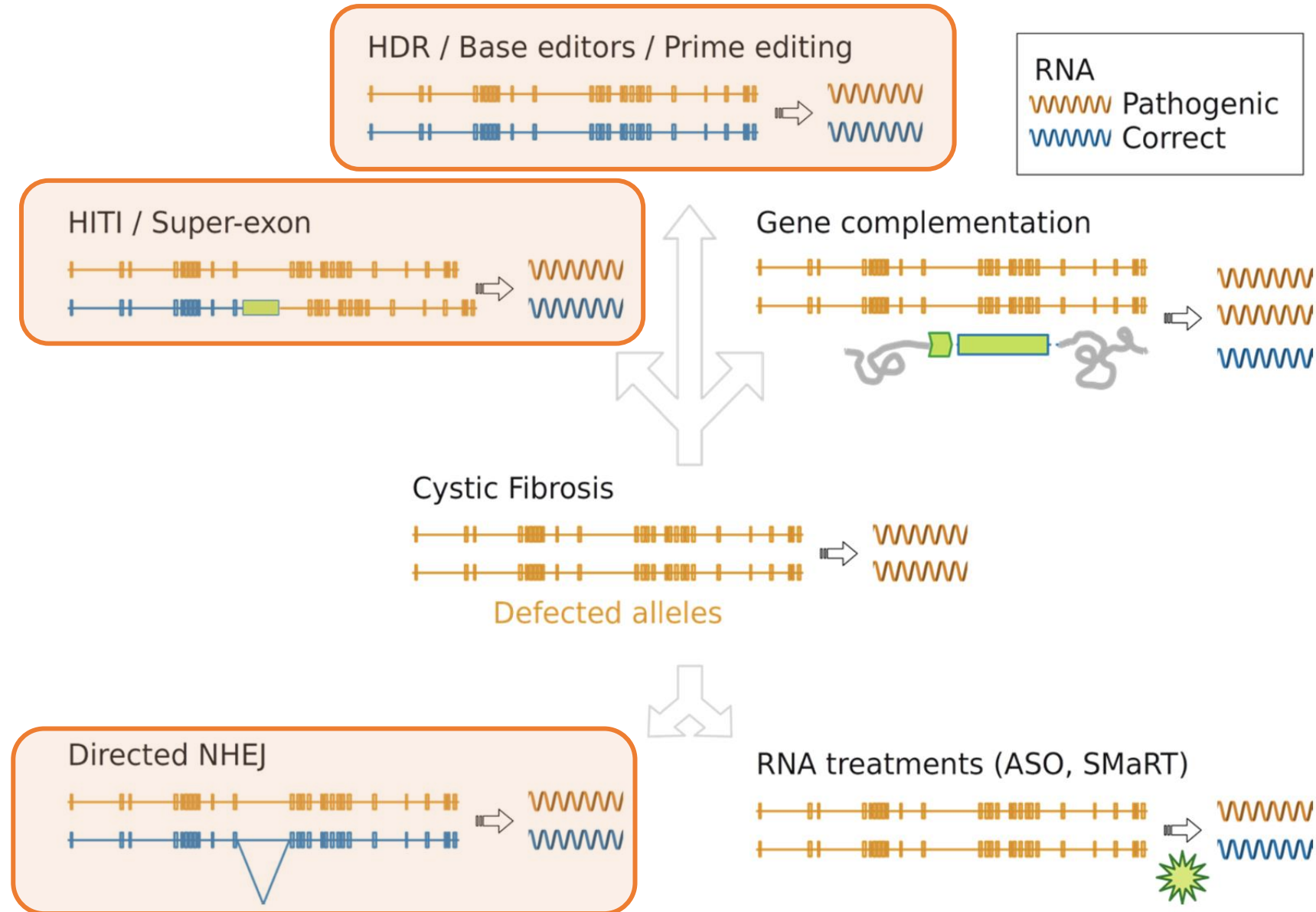
Viral methods to deliver genome editing cargo



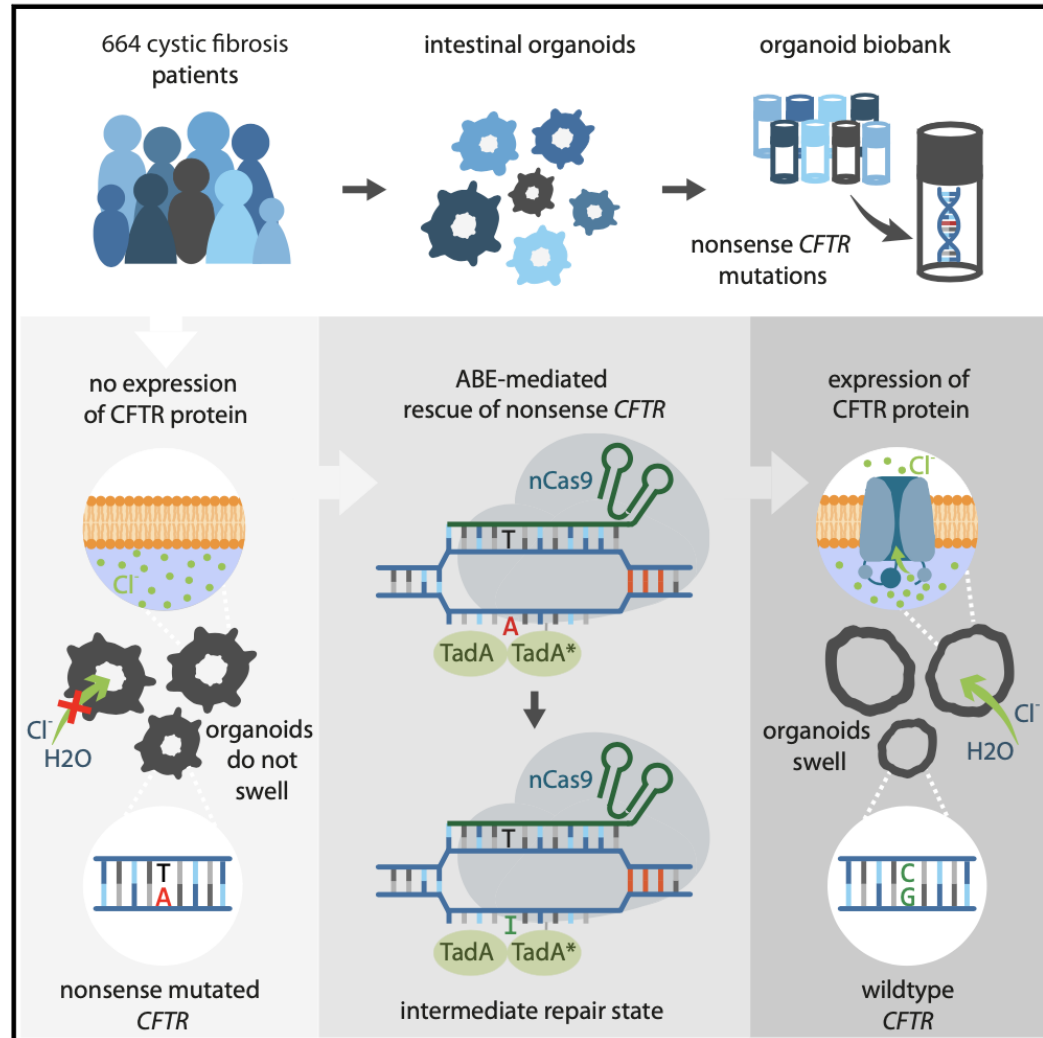
Non-viral methods to deliver genome editing cargo



Gene therapy for Cystic Fibrosis



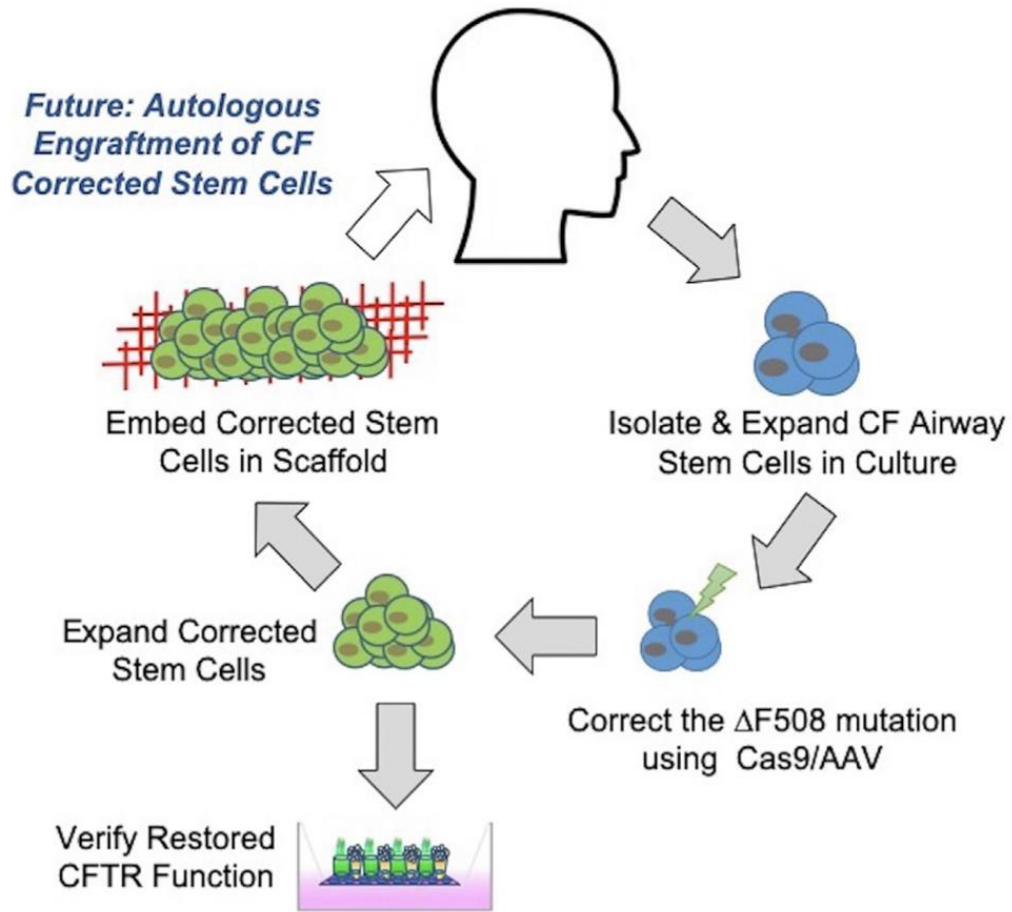
Second generation CRISPR-Cas tools: Base-editors and prime editors



CRISPR-Based Adenine Editors Correct Nonsense Mutations in a Cystic Fibrosis Organoid Biobank- Cell Stem Cells, 2020-Hans Clevers

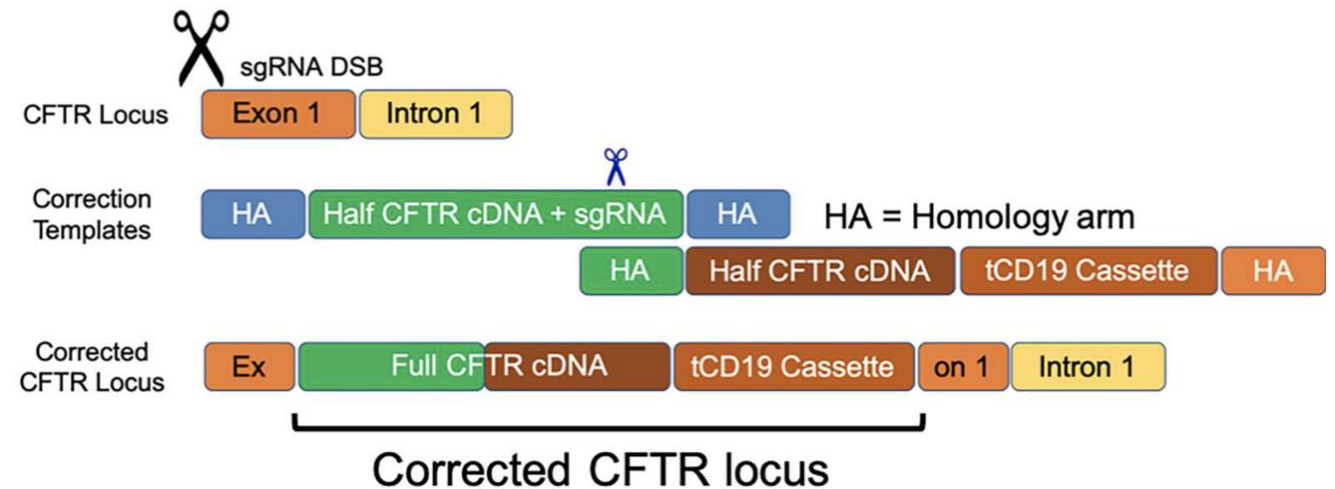
Ex-vivo gene replacement through CRISPR-Cas induced homologous recombination

Ex-Vivo Correction of Cystic Fibrosis (CF) Mutations in Airway Stem Cells



Targeted replacement of full-length CFTR in human airway stem cells by CRISPR/ Cas9 for pan-mutation correction in the endogenous locus – Mol Ther- 2021 M. Porteus

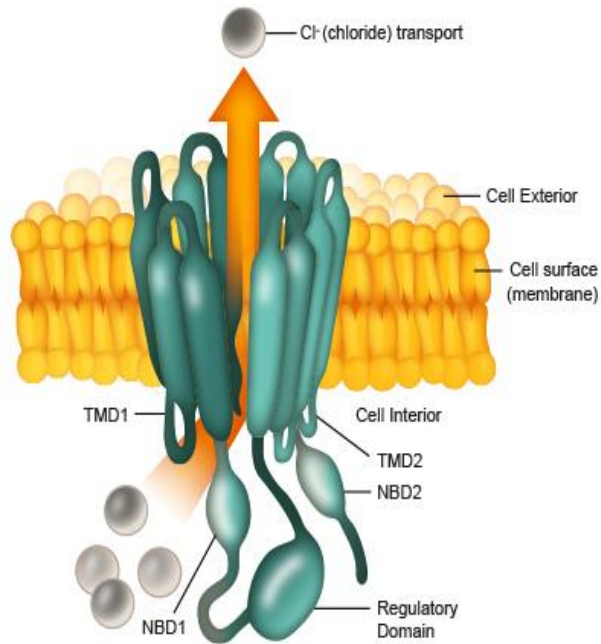
High-Efficiency, Selection-free Gene Repair in Airway Stem Cells from Cystic Fibrosis Patients Rescues CFTR Function in Differentiated Epithelia – Cell Stem Cell 2020 M Porteus



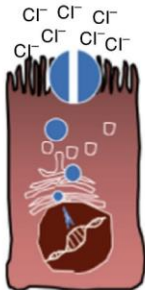


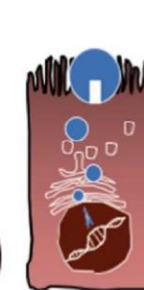
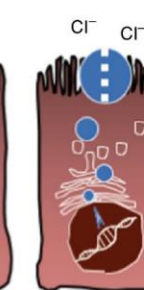

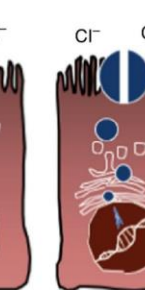
Correzione di una mutazione di splicing
3272-26A>G in CFTR tramite genome
editing con CRISPR-Cas

Cystic fibrosis

- autosomal recessive monogenetic disease caused by mutations in the **CFTR** gene
- ABC transporter-class ion channel that conducts chloride ions across epithelial cell membranes



**Cystic
Fibrosis**
**Transmembrane conductance
Regulator**

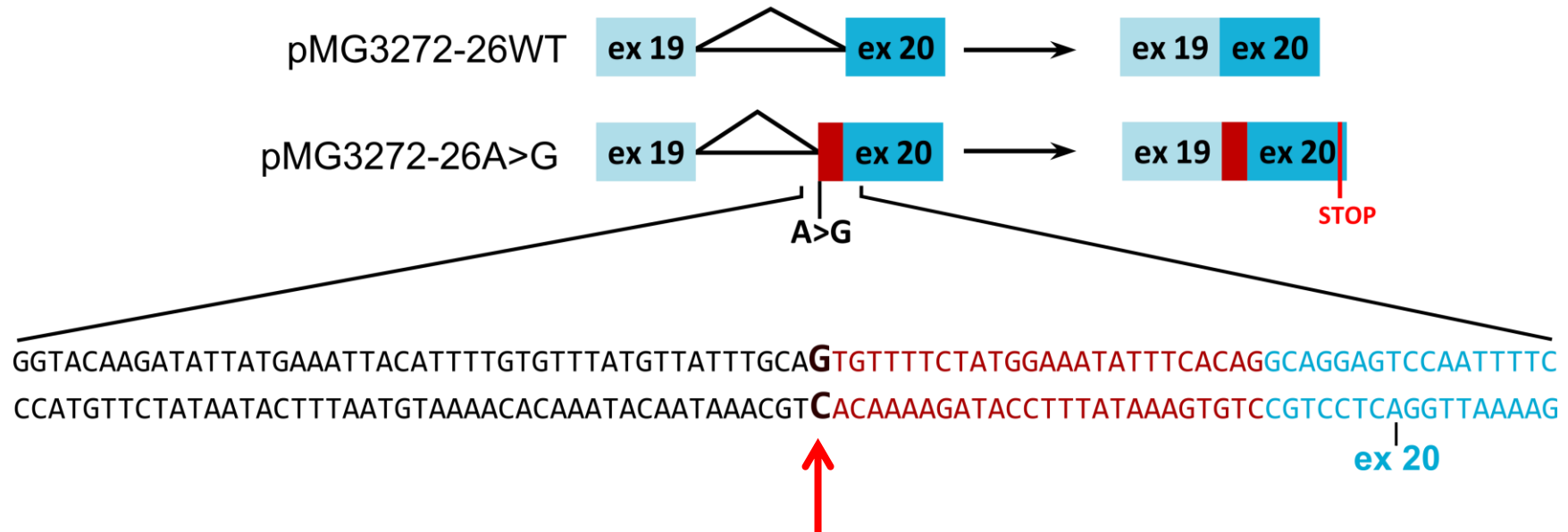
Class of mutation						
Normal	I	II	III	IV	V	VI
						
Molecular defect	No synthesis	Block in processing	Block in regulation	Reduced conductance	Reduced synthesis	Reduced half-life
Functional abnormality	Protein is not synthesized	Folding defect	Channel opening defect	Ion transport defect	Decreased protein synthesis	Decreased half-life of the protein
Main mutations	Gly542X Trp128X Arg553X 621+1G→T	Phe508del Asn1303Lys Ile507del Arg560Thr	Gly551Asp Gly178Arg Gly551Ser Ser549Asn	Arg117His Arg347Pro Arg117Cys Arg334Trp	3849+10kbC→T 2789+5G→A 3272-26A →G 5T	4326delTC Gln1412X 4279insA



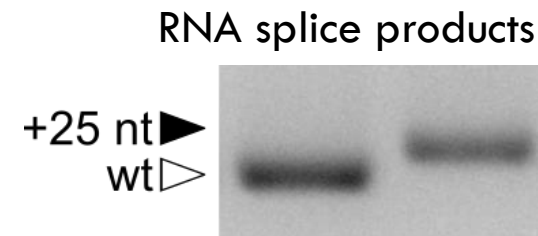
Splicing defects

3272-26A>G CF splicing mutation

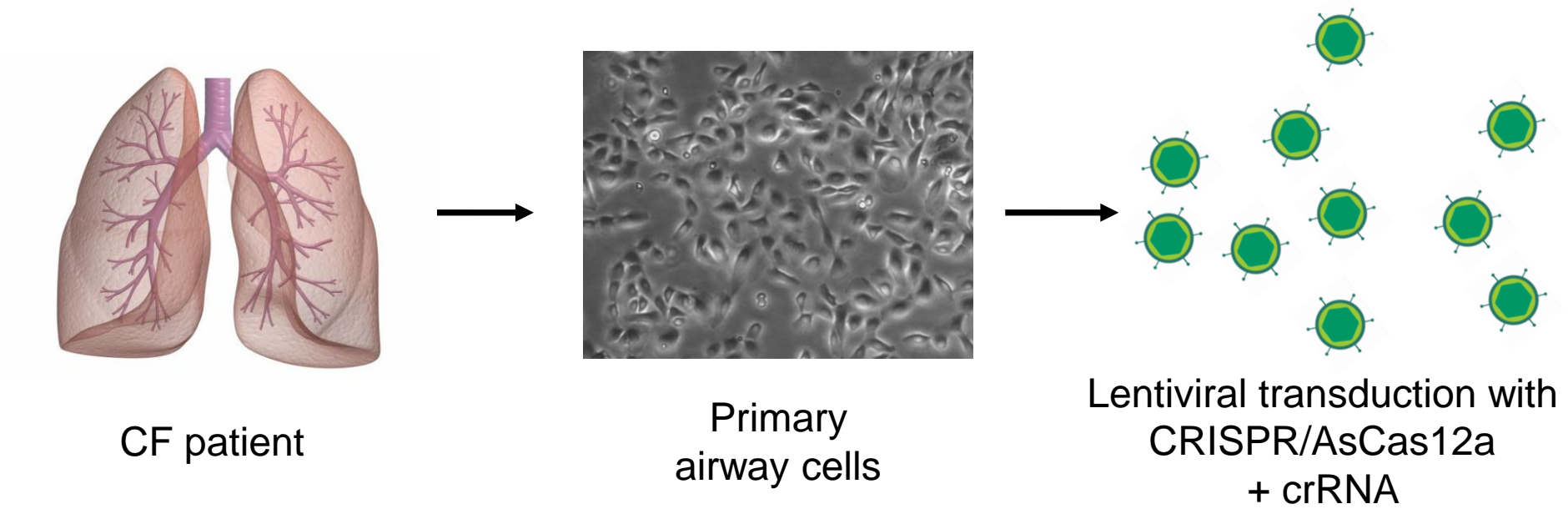
CFTR intron 19 MINIGENE models



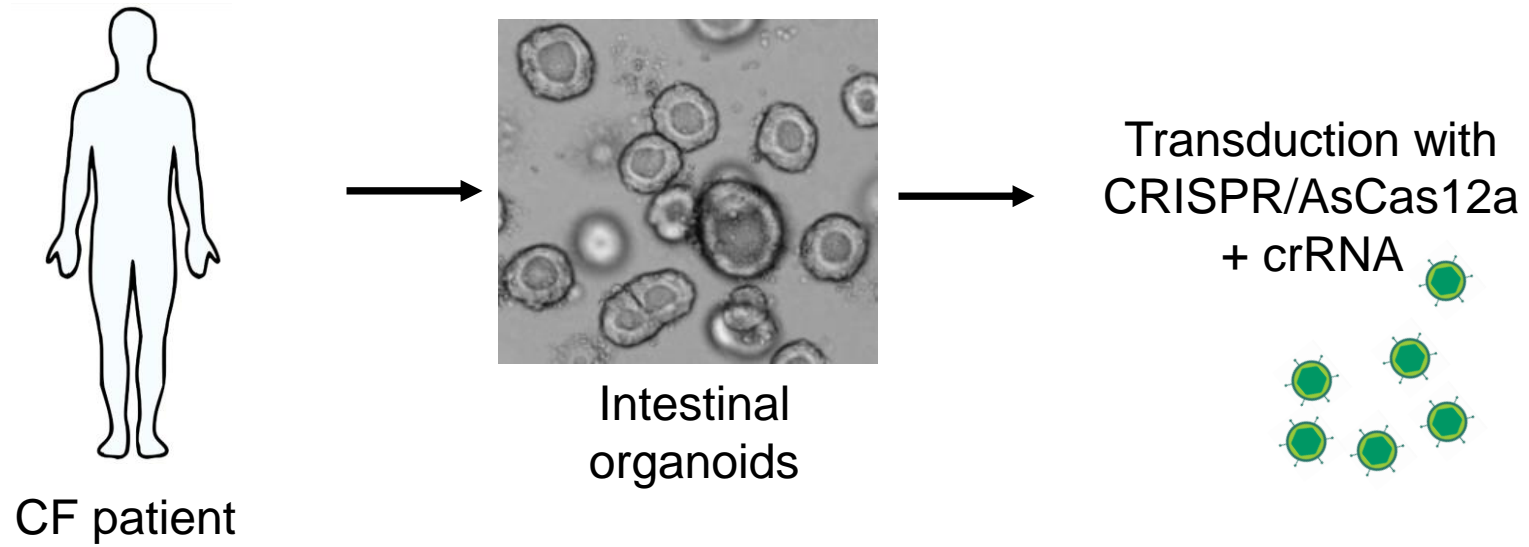
Alternative acceptor
splice site



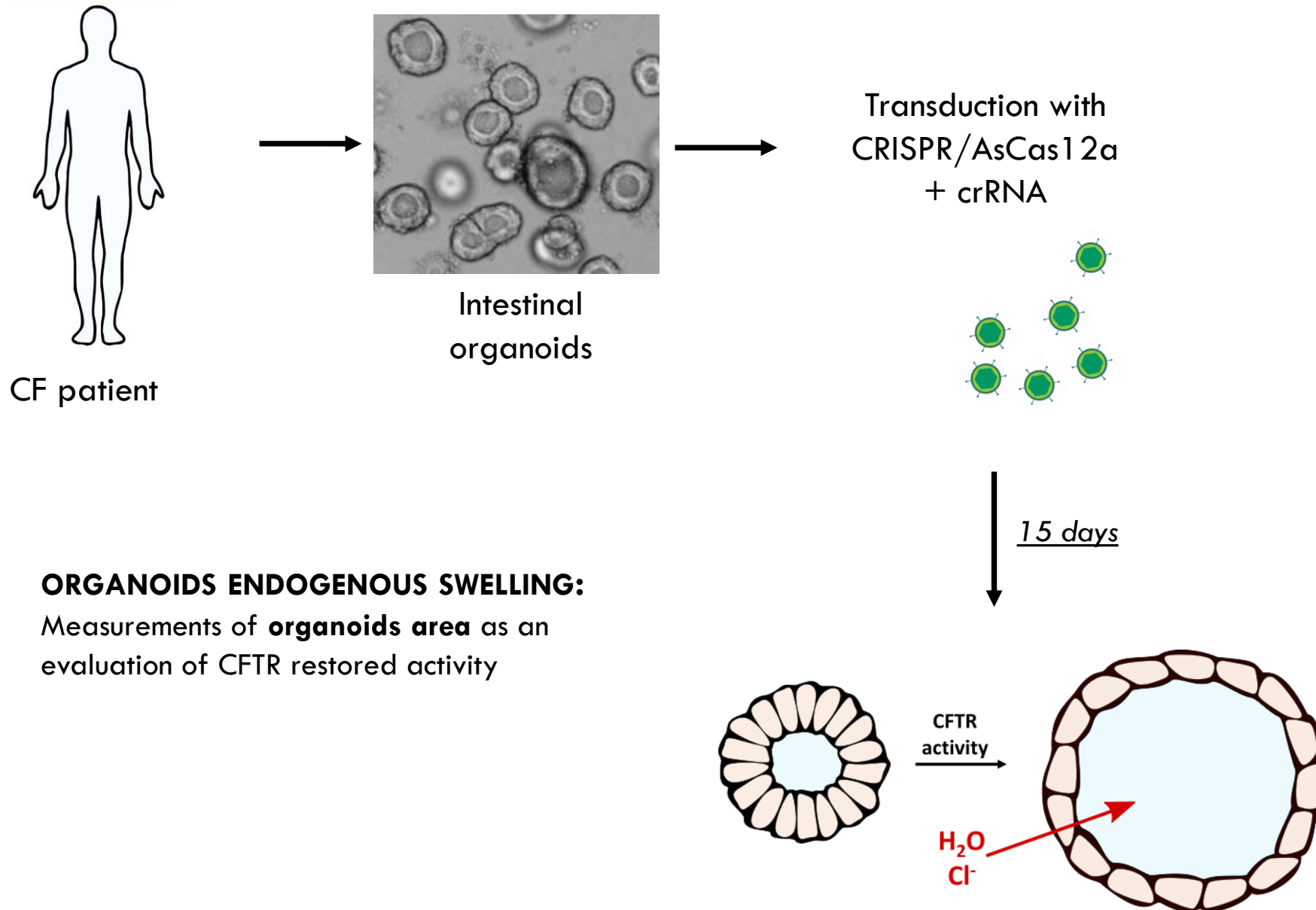
CFTR splicing repair in primary airway cells



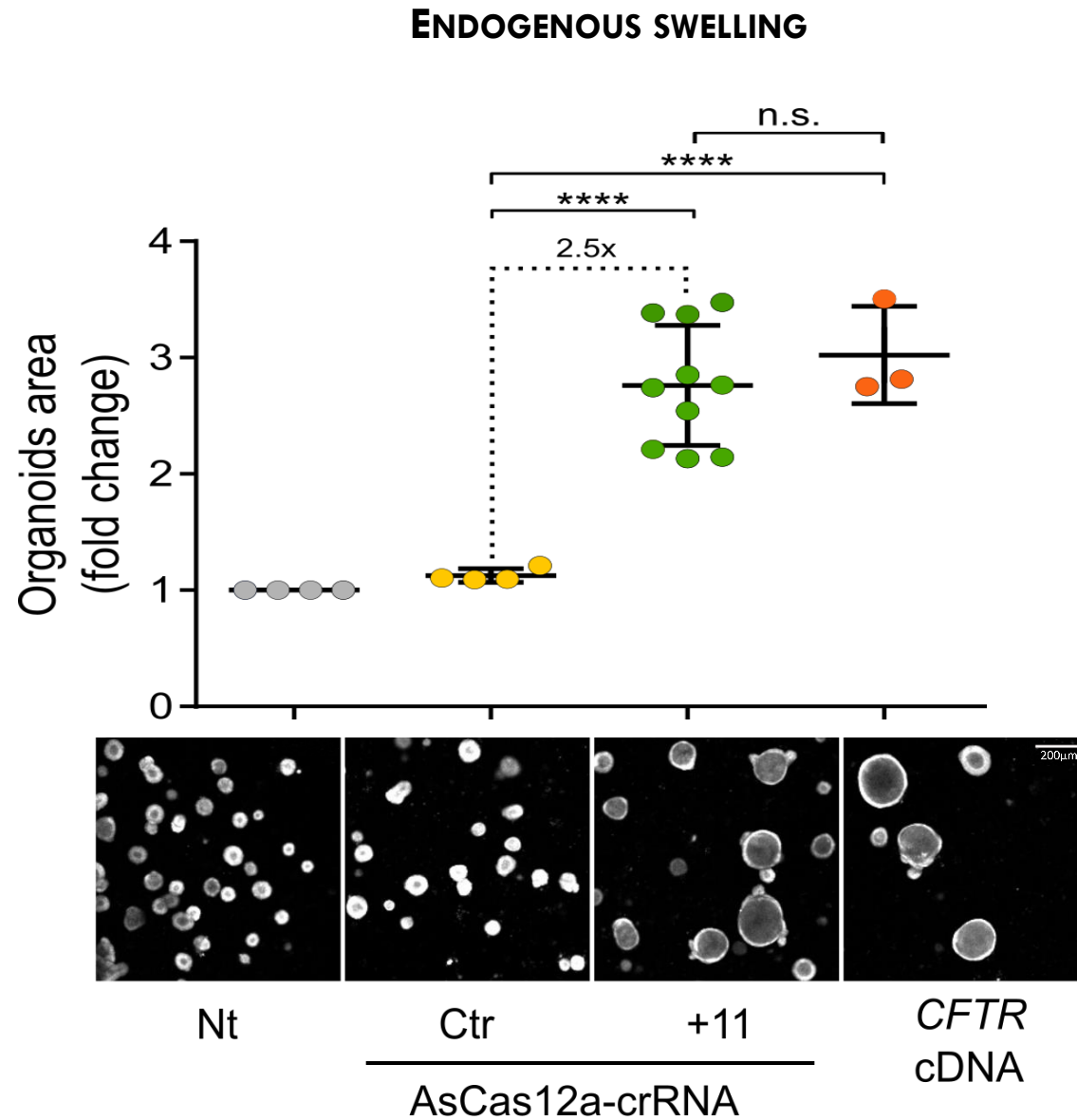
...and in intestinal organoids



Organoids experiments



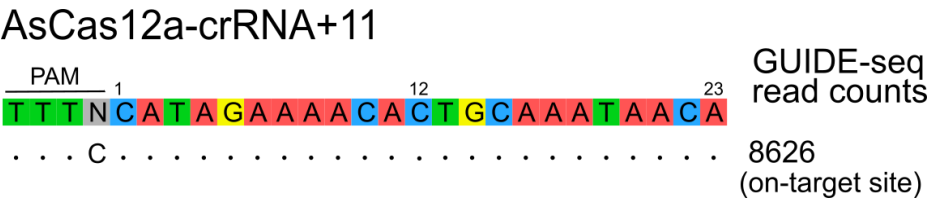
AsCas12a editing in 3242-26A>G organoids



**** $P < 0.0001$

crRNA OFF target analysis

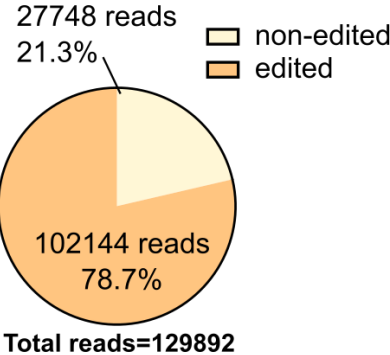
GUIDE-SEQ METHOD



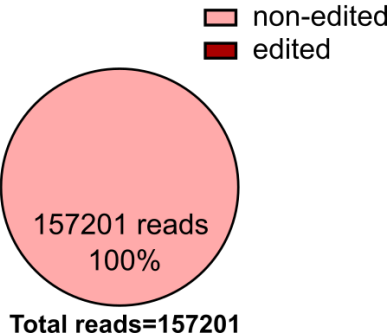
TARGETED DEEP-SEQUENCING

Primary airway cells

3272-26A>G allele

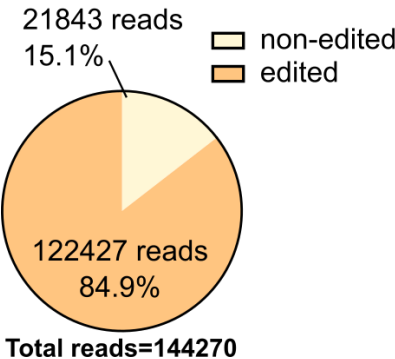


3272-26WT allele

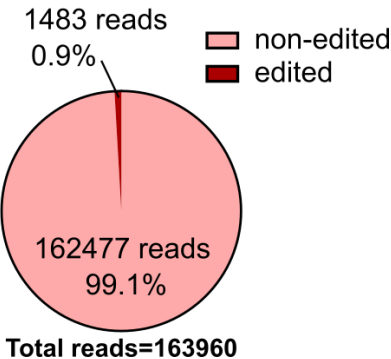


Organoids

3272-26A>G allele



3272-26WT allele



Genome editing per fibrosi cistica. Dove siamo:

La tecnologia CRISPR-Cas permette di correggere in-vitro la maggioranza (tutte) le mutazioni del gene *CFTR*.

Il successo in clinica della tecnologia dipende da:

1) DELIVERY

2) Precisione

3) Accessibilità del locus in cellule primarie (a seconda delle mutazioni)



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Acknowledgements

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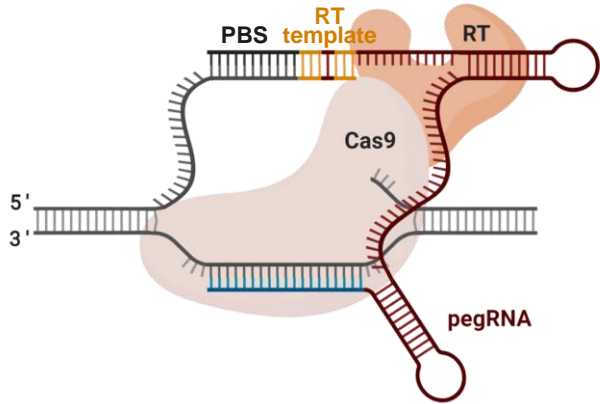
Marianne Carlon

Computational Metagenomics (UniTN)

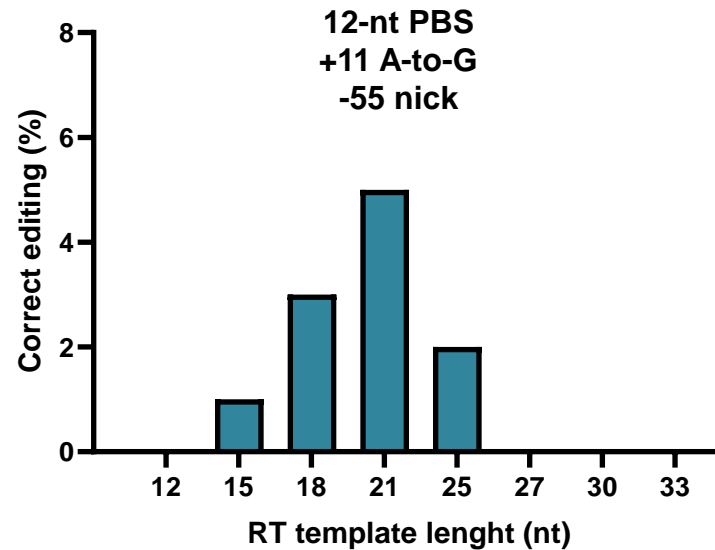
Nicola Segata



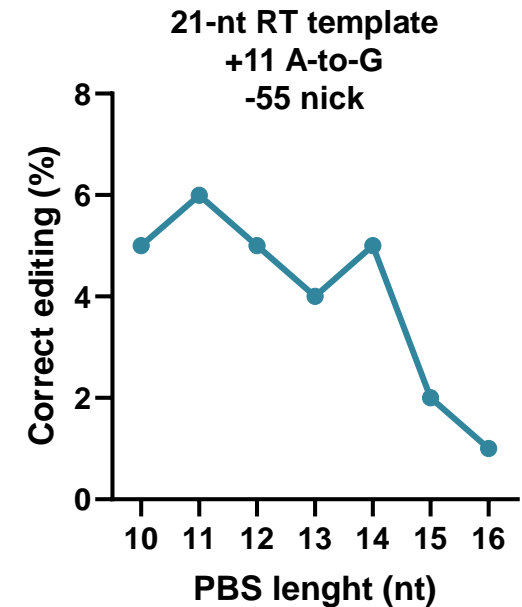
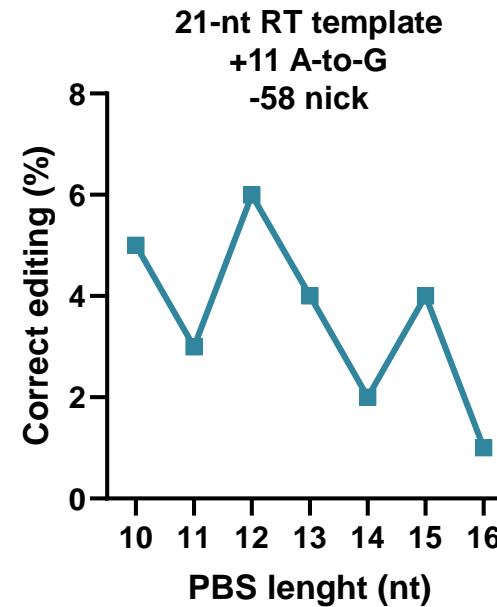
Prime editing: 2789+5G>A: pegRNA optimization



RT template length



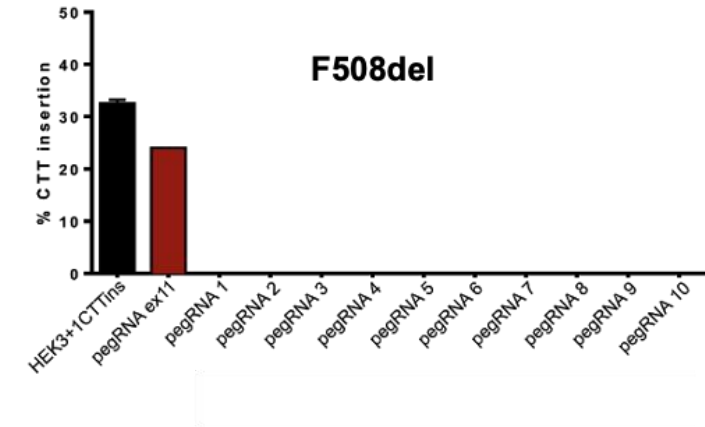
PBS length



A

pegRNAs	spacer sequence	3' extension	gRNA for PE3 strategy
HEK3 +1CTTins	GGCCCAGACTGAGCACGTGA	TCTGCCATCA AAG CGTGCTCAGTCTG	GTCAACCAGTATCCCGGTGC
pegRNA ex11	GGGAGAACTGGAGCCTTCAG	TTACCCCTCTGAAG AAG GCTCCAGTTC	TGGAGATGTCCTCTTCTAGT
pegRNA1	TCTGTATCTATATTCATCAT	T CTT TGGTGTTTCCTATGATGAATATAG	CATTCTGTTCTCAGTTTTCC
pegRNA2	TCTGTATCTATATTCATCAT	T CTT TGGTGTTTCCTATGATGAATATAGAT	
pegRNA3	TCTGTATCTATATTCATCAT	T CTT TGGTGTTTCCTATGATGAATATAGATACA	
pegRNA4	TCTGTATCTATATTCATCAT	T CTT TGGTGTTTCCTATGATGAATATAGATACAGA	
pegRNA5	ACCATTAAAGAAAATATCAT	AGGAAACACCA AAG ATGATATTTTCTT	TGGAGATGTCCTCTTCTAGT
pegRNA6	ACCATTAAAGAAAATATCAT	AGGAAACACCA AAG ATGATATTTTCTTTA	
pegRNA7	ACCATTAAAGAAAATATCAT	AGGAAACACCA AAG ATGATATTTTCTTTAAT	
pegRNA8	ACCATTAAAGAAAATATCAT	AGGAAACACCA AAG ATGATATTTTCTTTAATGGT	
pegRNA9	CAGTTTTCTGGATTATGCC	ACCA AAG ATGATATTTTCTTTAATGGTGCCAGGCATAATCCAGGAAAAC	TGGAGATGTCCTCTTCTAGT
pegRNA10	CAGTTTTCTGGATTATGCC	ACCA AAG ATGATATTTTCTTTAATGGTGCCAGGCATAATCCAGGAAAAC	

B



Use of CRISPR in biomedical field



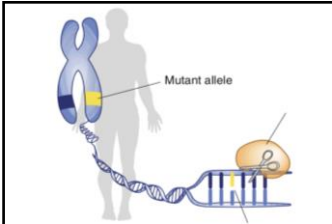
Life science research

Genome editing for gene ko, screenings (drug), cell marking etc



Animal model

Disease model to understand pathogenic mechanisms and therapeutic development.



Gene therapy

Not just gene complementation but genes repair or ko of gain of function mutations



Embryo genome editing

Treatments of genetic disease



Virus detection and control

Identification and cleavage of viral nucleic acids or neutralization of viral co-factors (e.g. CCR5)

Genome editing
TECHNOLOGY

Genome editing
APPLICATIONS/CLINIC

SLOW LANE

Next **LTS** in 3 months

FAST LANE

Latest **Release** available



Use of CRISPR



De-extinction

Woolly mammoths (*Mammuthus primigenius*) to extinction.



Disease control

Disease resistance is one of the most popular applications for CRISPR help to stem the dramatic loss of honeybees.



Better food production

The US Food and Drug Administration approved the first transgenic animals for human consumption: fast-growing salmon.



Making drugs

chickens with components required for CRISPR integrated directly into their genomes (CRISPR chickens) to edit chicken DNA: 'farmaceuticals' — drugs created using domesticated animals.



Vector control

genetically modifying mosquitos to prevent the spread of diseases such as dengue or malaria.

Take-home message

Genome editing for cystic fibrosis:

Technological opportunities through CRISPR-Cas:
the majority/all mutations can be corrected this technology

More advancements needed:

1) DELIVERY

2) Precision

3) Genomic locus accessibility

Genome-editing in the literature

